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## Determination of ultra-trace amount Chromium in water samples with 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH) by the potassium chlorate ion in sulfuric acid with kinetic spectrophotometric method

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### ABSTRACT

A new kinetic spectrophotometric method for the determination of trace amount Chromium(III) in Tea real samples has been described based on its catalytic effect on the oxidations of 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), by potassium chlorate in sulfuric acid. The reaction is followed spectrophotometrically by measuring the decrease in the absorbance at 469 nm. Under the optimum conditions of 0.2 molL<sup>-1</sup> sulfuric acid, 1.0 × 10<sup>-4</sup> molL<sup>-1</sup>, 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), 0.01 mol L<sup>-1</sup> potassium chlorate at 35°C, calibration graph in the rang of 0.001-0.06 mg L<sup>-1</sup> Chromium(III). Concentration was obtained with detection limit of 0.2303 g mL<sup>-1</sup> by the fixed-time method of 5.5 min. The relative standard deviation for 0.05 mg L<sup>-1</sup> Chromium(III) is %90. No serious interference was identified. The applicability of the method was demonstrated by the determination of the Chromium(III) in Water samples.

**Keywords:** Chromium(III); Kinetic; 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH); Determination; Spectrophotometric.

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### INTRODUCTION

Chromium is a major water pollutant, usually as a result of some industrial pollution including tanning factories, steel work, dyeing or chromium plating, wood preservation, and artificial fertilizers[1]. For this reason, determination ion of chromium in environmental samples and natural water, waste water samples and soil samples has become very important. The USEPA has set concentration 0.1ng/L of total chromium for drinking water as “maximum contaminant level goals” WHO states that the guideline value of 50µg/L of chromium (VI) because of these importance of chromium species, their accurate and facile determinations are in important in the chemistry[2]. The sophisticated techniques, such as inductively coupled plasma- mass spectrometry (ICP-MS), inductively coupled plasma-atomic emission spectrometry (ICP-AES), electrochemical analysis, spectrophotometry, neutron activation analysis and atomic absorption spectrophotometry (AAS), been adopted for sensitive assays for both oxidation states of chromium. These methods have disadvantage of cost and instruments used in regular analysis. AAS often has low sensitivity due to the matrix effect of samples such as salinity[3]. environmental samples at low levels and has matrix effects with detection limit of lead[4]. The traditional methods of spectrophotometers are employed with separation from matrix elements. Extractive and micellar sensitized spectrophotometer with ability to separate and preconcentrat of chromium solve these problems and lead to a higher confidence level and easy determination of the trace elements by less sensitive, but more accessible instrumentation such as spectrophotometer. For this purpose various preconcentration method of chromium in micellar media are used[5]. The spectrophotometric and extraction method based on co-ordination with different ligands have been described for chromium determination[6-9]. Hence, sensitive and selective methods for determination of Chromium in foodstuffs, drinking waters and drinks are of great interes. There have been numerous spectrophotometric methods for the

determination of Chromium. The oxidation of Chromium to permanganate and a measurement of the absorption of the charge-transfer band of permanganate at 558 nm has long been used as a standard method for Chromium determination this method suffers from low sensitivity. Spectrophotometric methods based on complex formation with chromogenic reagents provide good sensitivity, but have showed serious interferences from many cations and anions despite the high detection limit of  $0.05 \text{ ng mL}^{-1}$  [10,11]. Numerous kinetic methods have been reported based on the catalytic effect of  $\text{Cr}^{3+}$  on the oxidation of organic compounds with suitable oxidants. Although these methods have shown good sensitivity, they are time consuming and irreproducible, since it is difficult to control the timing of the reaction, which is variable from one experiment to another [12– 16], Some recent enhancements of the method using a flow injection technique have been reported [17 – 19], However, if the reaction time is longer than the flow rate, the method will not be efficient and still irreproducible. Several efforts in crease the reaction rate by temperature have been reported; however, either the sample degrades or airbubbles will develop that hinder the flow rate [20-26]. In this work, the catalytic effect of Chromium(III) on the oxidation of potassium chlorate in the presence of 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), was investigated. A catalytic kinetic Spectrophotometric method for the determination of Chromium(III) was developed [27,28]. The proposed method is extremely sensitive, with higher selectivity and is a simpler procedure than the three methods mentioned above. The method was applied successfully to the determination of total Chromium(III) in Water samples.

## MATERIALS AND METHODS

### 2.1. Apparatus

All the determinations of the analytes were carried out using a Sens AA GBC double beam atomic absorption spectrometer (AAS) equipped with deuterium background corrector. Hollow cathode lamps were used as radiation sources and the operational conditions of the equipment were established according to the manufacturer's recommendations for each element. An adjustable capillary nebulizer and supplies of acetylene and air were used for the generation of aerosols and atomizations. The UV/Vis spectra were obtained from a Perkin-Elmer, model Lambda 2 spectrophotometer. A Genway model 3510 pH/Ion meter with a combined glass electrode was used for pH measurements. Laboratory glassware was kept overnight in 10% nitric acid solution. A NBE ultrathermostat (VEB Prufgerate – WerkMedingen, Germany) was used to control the temperature.

### 2.2. Reagents and materials

All chemicals, except barium di phenyl aminosulfonate, were prepared of the analytical grade purchased from Merck Company, solution (Merck, Darmstadt, Germany). The solutions were prepared with doubly distilled water. The concentrations of the stock solutions were as follows: 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH),  $1.0 \times 10^{-4} \text{ mol L}^{-1}$ , potassium chlorate,  $0.1 \text{ mol L}^{-1}$  sulfuric acid,  $0.2 \text{ mol L}^{-1}$  Chromium(III) sulphate, 1000.

### 2.3. Pretreatment of real samples

#### Water samples

Analysis of water samples for determination of Cr(III) ion content was performed as following: 200 mL of sample was poured in a beaker and 8 ml concentrated  $\text{HNO}_3$  and 3 ml of  $\text{H}_2\text{O}_2$  of (30%) for elimination and decomposition of organic compound were added. The samples, while stirring was heated to one tenth volume. After adjustment of samples pH to desired value the were performed according to general described procedure [29].

### 2.4. Procedure

A typical kinetic Spectrophotometric method experiment required the following steps: 1 ml of manganese solution equated to 0.01 ml grams was added to the 10 ml volumetric flask. Than 1 ml gram of potassium chlorate ( $0.1 \text{ mol L}^{-1}$ ) and 1 ml of sulfuric acid solution ( $0.2 \text{ mol L}^{-1}$ ) was added. By adding the first drop of 1 ml 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH)  $1.0 \times 10^{-4} \text{ mol L}^{-1}$ , to the volumetric flask, the time of the reaction beginning is recorded, after 5 sec the solution is mixed for 30 sec, and then it is volume by adding the distilled water, a sufficient amount of the solution was added to a 1 cm cell, the difference between the quantities of the absorption in a wavelength equal to 469 nm in a time interval equal to 1-5.5 min was measured by mean of spectrophotometer (A As). All these steps would be repeated for a non catalytic reaction without the presence of manganese as the catalyst element (AAb), finally (AA)AAbblank-AAsample is calculated. 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), oxidation is traced in the acidic medium by potassium chlorate, which its wavelength is 469 nm. The absorption spectra in an aqueous solution are shown in figures 1 and 2.

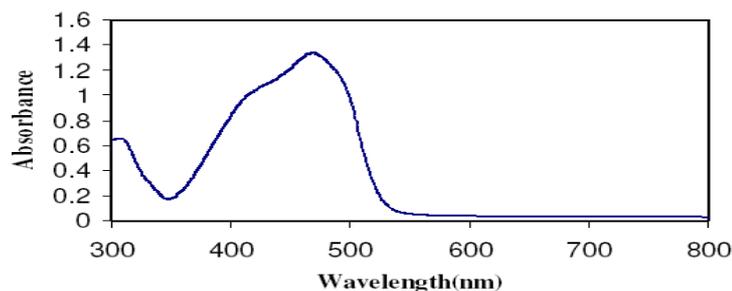


Figure 1: The absorption spectra oxidation of product 2- droxynaphthaldehydebenzoylhydrazone

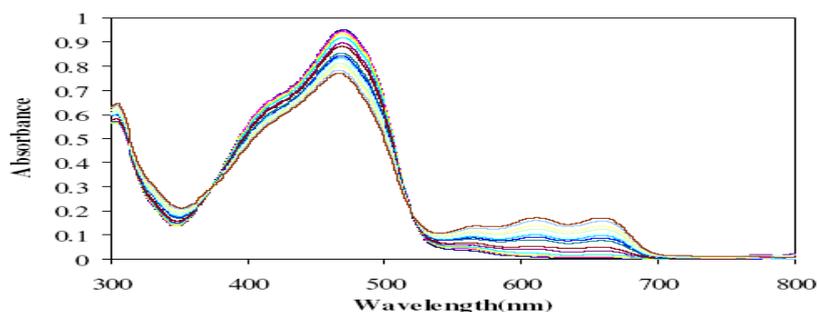


Figure 2: The absorption spectra oxidation of product 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), 20min mg L<sup>-1</sup>

## RESULTS AND DISCUSSION

### 3.1. Investigation of ligand complexation with metal ions

The complexation studies between the ligand and Cr(III) ion were carried out to determine the stoichiometry of the metals - ligand reactions as well as the formation constants and the molar absorptivities of these complexes. The stoichiometry between the ligand and each of the metal ions were determined from the absorbance-mole ratio data. The plots at 469 nm (Figure 3), clearly show a 1:1 (ligand to metal) stoichiometry for all the metal complexes. The formation constants,  $K_f$ , and the molar absorptivities,  $\epsilon_{ML}$ , of the resulting complexes were then evaluated from the absorbance-mole ratio data and with the aid of a nonlinear curve fitting of the theoretical absorbance,  $A_{Theo}$ , to the experimental absorbance,  $A_{Exp}$ , using Microsoft Excel Solver program[30]. When known concentrations of metal,  $C_M$ , is mixed with a constant concentration of ligand and ML complexes are formed, the following equations can be written;

$$\begin{aligned}
 L + M &\leftrightarrow ML & K_f &= [ML] / [L] [M] \\
 [L] &= C_L - C_M + x & [M] &= x \quad [ML] = C_M - x \\
 K_f x^2 + (K_f C_L - K_f C_M + 1) x - C_M &= 0 \\
 A_{Theo} &= A_L + A_{ML} = (A_0 / C_L) [L] \\
 &+ \epsilon_{ML} K_f C_M [L] / (1 + K_f [L]) \quad (2)
 \end{aligned}$$

where  $A_0$  is the initial absorbance of the ligand with concentration of  $C_L$  before any metal ions addition. Also,  $x$  is a reversed amount of ligand or metal reproduced from decomposition of ML at equilibrium which is equal to  $[M]$  value under  $C_M \geq C_L$  condition. The procedure actually consists of two mutual steps. First, by initial guessing for  $K_f$  and solving Equation (1),  $x$  value is obtained which yields  $[L]$  parameter from  $[L] = C_L - C_M + x$  then  $A_{Theo}$  is calculated by the substitution of  $[L]$  in Equation (2) ( $C_L$ ,  $C_M$  and  $A_0$  are known parameters). In the second step, for fitting the curve of  $A_{Theo}$  to that of  $A_{Exp}$ , minimization of SSR, sum of squared of residual, is performed by the Excel Solver program and two adjustable parameters  $K_f$  and  $\epsilon_{ML}$  are found. The results, which are summarized in (Table 1), show the values of formation constants for the considered metal complexes.

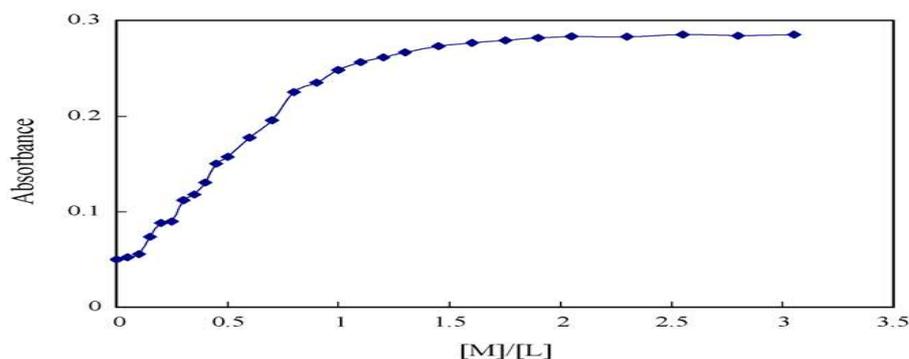


Figure3. Absorbance-mole ratio plot for Cr- (HNPBH) complexes at  $[M] = 469 \text{ nm}$ ,  $[\text{HNPBH}] = 1.0 \times 10^{-4} \text{ mol/L}$  in Methanol

Table 1. Formation constants and molar absorptivities of the metal complexes

Type of complex	$K_f$	$\epsilon_{ML}$	SSR <sup>a</sup>
Cr- (HNPBH)	$1.02 \times 10^{-4}$	$2.71 \times 10^{-3}$	$1.22 \times 10^{-3}$

$$^a\text{SSR} = \sum (A_{\text{Exp}} - A_{\text{Theo}})^2$$

### 3.2. Effect of time on the reaction rate

As it was expressed in the method, to obtain optimum time of the reaction, 1 ml Cr(III)  $0.01 \text{ mg L}^{-1}$  solution, 1 ml potassium chlorate  $0.1 \text{ mol L}^{-1}$ , 1 ml sulfuric acid solution  $0.2 \text{ mol L}^{-1}$  and 1 ml 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH),  $1.0 \times 10^{-4} \text{ mol L}^{-1}$  are added to the volumetric flask 10 ml and by adding distilled water. Absorption of so lutions was measured in the 0-11 min interval of time. The above mentioned operation was repeated for biank solution ( the solution without Cr(III)). Changes in absorption based on the time at 30 centigrade degrees temperature are shown in figure 4 . 5.5 min was selected as the optimum time.

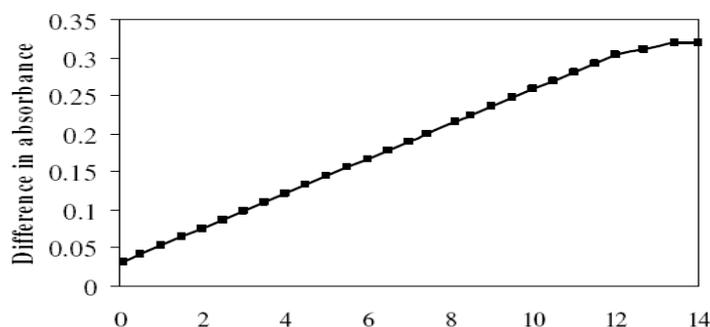


Figure 4: The effect of time on the reaction rate

### 3.3. Effect of sulfuric acid concentration on the rate of reaction

As it was expressed in the method, to obtain optimum sulfuric acid concentration of the reaction, 1 ml Cr(III)  $\text{mgL}^{-1}$  solution, 1 ml potassium chlorate  $0.2 \text{ mol L}^{-1}$ , 1 ml sulfuric acid solution  $0.1$  to  $10 \text{ mol L}^{-1}$  and 1 ml, 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH),  $1.0 \times 10^{-4} \text{ mol L}^{-1}$  are added to the volumetric flask 10 ml and by adding distilled water. Absorption of solution was measured after 5.5 min. The above mentioned operation was repeated for biank solution ( the solution without Cr(III) ). As it is shown in figure 5, the 0.2 molar concentration of sulfuric acid has the most changes in the absorption, so that, it was selected as the optimum concentration of the acid.

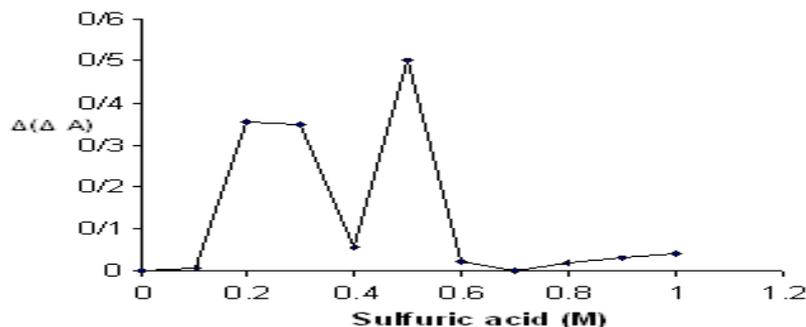


Figure 5: The effect of sulfuric acid concentration on the rate of reaction

### 3.4. Effect of 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), on the reaction rate

To inspecting The effect of 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), on the reaction rate, 1 ml Cr(III)  $0.01 \text{ mg L}^{-1}$  solution, 1 ml potassium chlorate  $0.1 \text{ mol L}^{-1}$ , 1 ml sulfuric acid solution  $0.2 \text{ mol L}^{-1}$  and 1 ml, 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH),  $1.56 \times 10^{-5}$  to  $2.5 \times 10^{-3} \text{ mol L}^{-1}$  are added to the volumetric flask 10 ml and by adding distilled water. Absorption of solution was measured after 5.5 min. The above mentioned operation was repeated for blank solution (the solution without Cr(III)). The results are shown in and figure 5, based on those results  $1.0 \times 10^{-4} \text{ mol L}^{-1}$  was selected as the desired concentration.

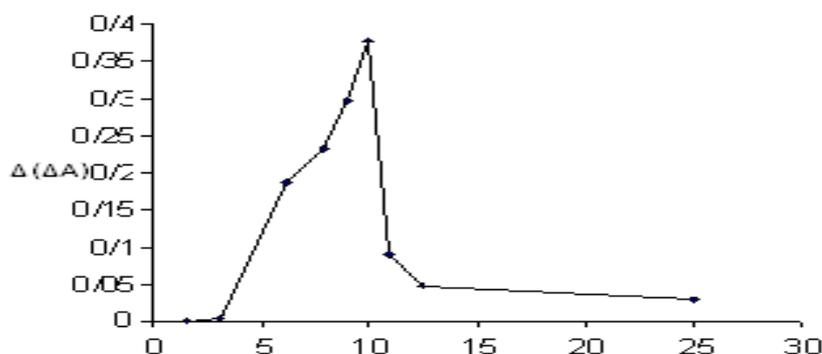


Figure 5: The effect of 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), on the reaction rate

### 3.5. Effect of temperature on the reaction rate

At first put the cells including: Cr(III), potassium chlorate, 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH), distilled water and volumetric 10 ml flasks in the thermostat to reach to the desired temperature. After they go to the equilibrium temperature, 1 ml Cr(III)  $0.01 \text{ mg L}^{-1}$  solution, 1 ml sulfuric acid solution  $0.2 \text{ mol L}^{-1}$  and 1 ml 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH),  $1.0 \times 10^{-4} \text{ mol L}^{-1}$  are added to the volumetric flask 10 ml and by adding distilled water. Absorption of solution was measured after 5.5 min. The above mentioned operation was repeated for blank solution ( the solution without Cr(III)). As it is shown in figure 6, 35 centigrade degree was selected as the desired temperature.

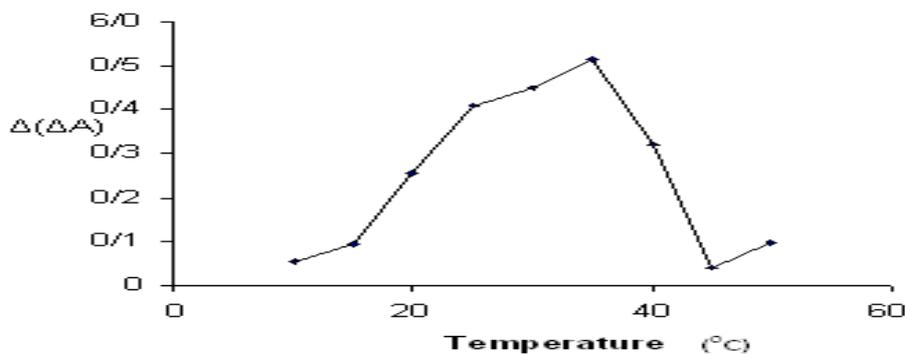


Figure 6: The effect of temperature on the reaction rate

### 3.6. Effect of ionic power of the medium was also

inspected; potassium chlorate and potassium nitrate were used for this purpose. 1 ml Cr(III)  $0.01 \text{ mg l}^{-1}$  solution, 1 ml potassium chlorate  $0.1 \text{ mol L}^{-1}$ , 1 ml sulfuric acid solution  $0.2 \text{ mol L}^{-1}$  and 1 ml 2-hydroxynaphthaldehydebenzoylhydrazone (HNPBH),  $1.0 \times 10^{-4} \text{ mol L}^{-1}$  are added to the volumetric flask 10 ml and by adding distilled water. Absorption of solution was measured after 5.5 min. as we can see in figure 7 results show that the effect of ionic power on the reaction rate is neglectible, and can be ignored.

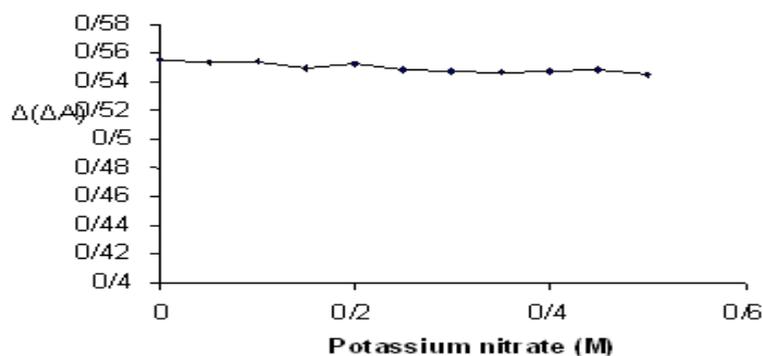


Figure 7: The effect of Ionic power on the reaction rate

### 3.7. Calibration graph and reproducibility

Reaching to the standard curve of Cr(III) in the reaction, the system went into the desired condition. So different volumes of Cr(III) ( $100 \text{ mg L}^{-1}$ ), 1 ml potassium chlorate  $0.1 \text{ mol L}^{-1}$ , 1 ml sulfuric acid solution  $0.2 \text{ mol L}^{-1}$  and 1 ml 2-hydroxynaphthaldehydebzoylhydrazone (HNPBH),  $1.0 \times 10^{-4} \text{ mol L}^{-1}$  are added to the volumetric flask 10 ml and by adding distilled water. Absorption of solution was measured after 5.5 min. the above mentioned operation was repeated for blank solutions. The examination/test was done at 35 centigrade degrees. Results are shown figure 8.

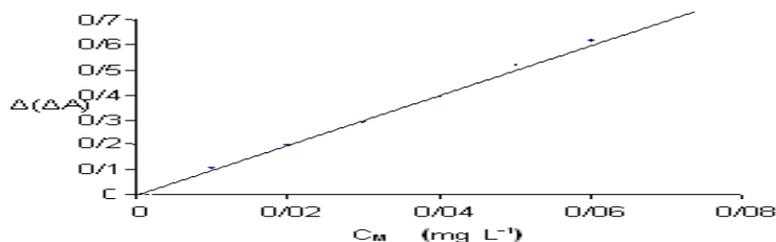


Figure 8: Calibration graph for Cr(III)

### 3.8. Effect of foreign ions

The accuracy and reliability of this method was evaluated by comparing the results obtained for the same samples by an AAS method. The results estimated by AAS and spectrophotometry are shown in Table 4. The results estimated by spectrophotometry are much less as compare to results obtain by AAS. It may be attributed to the interference of foreign ions in the determination of chromium. The interfering ions. These ions may interfere with chromium(III). In order to assess the potential analytical applications of the proposed kinetic reaction, the influence of foreign ions on the determination of nickel was investigated[31-33]. The tolerated limits for the ions assayed are shown in Table2, (was relative errors less than 5%). As can be seen, the proposed method is highly selective.

Table 2. Effects of the matrix ions on the recoveries of the examined Chromium(III) ion (N=6)

Ion	Added As	Tolerance Limit, $\text{mg L}^{-1}$
Cl <sup>-</sup> , K <sup>+</sup> , Na <sup>+</sup>	KCl, NaCl	1000
Mg <sup>2+</sup> , Ca <sup>2+</sup>	chloride salts	800
Cu <sup>2+</sup>	Nitrate salts	700
HCO <sub>3</sub> <sup>-</sup>	NaHCO <sub>3</sub>	1000
SCN <sup>-</sup>	KSCN	1000
PO <sub>4</sub> <sup>3-</sup>	Na <sub>3</sub> PO <sub>4</sub>	400
Fe <sup>3+</sup>	Nitrate salts	300
CO <sub>3</sub> <sup>2-</sup>	Na <sub>2</sub> CO <sub>3</sub>	500
Mn <sup>2+</sup>	Nitrate salts	150
Cd <sup>2+</sup>	Nitrate salts	250

### 3.9. Application

To assess the applicability of the method to real samples with different matrices, containing various amounts of diverse ions were used. For accuracy and reliability of proposed method, spiking experiments and independent analysis were used. The proposed method was applied to the determination of total Chromium(III) in water samples. In Table3 the results obtained are shown, and compared with those obtained by atomic absorption spectrophotometry. The level of the analyte ions were found below the detection limit of related element. The results of replicate three analyses of each sample show that the ions recoveries are almost quantitative with a low RSD. The

recovery of spiked samples is satisfactory reasonable and was confirmed using standard addition method, which indicate the capability of the proposed method for the determination of trace amounts of these elements in different samples.

**Table 3. Recovery of trace Chromium (III) from water sample after application of presented procedure (N=6)**

sample	Added (ng L <sup>-1</sup> )	Founded (ng L <sup>-1</sup> )	RSD %	Recovery %
Tap water	0	103.3	1.1	---
	100	161.9	1.8	103
River water	0	61.8	1.1	---
	100	164.3	1.3	95.3

### CONCLUSION

The proposed method is very simple, highly selective, sensitive and reproducible for the determination of Chromium(III). The method also exploits low-cost instrumentation and overcomes the problems associated with previously reported spectrophotometric method for the determination of Chromium[34-37]. The method is simple, accurate can be applied for the determination of analytes in environmental samples [38,39].

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### REFERENCES

- [1] Office of drinking water. Health advisory chromium Washington DC, US. Environmental Protection Agency. **1987**.
- [2] P. Rashida, A. Maria, M. Summayia, P. Abida, *Journal of Pharmaceutical and Chemical Sciences*, 2(1), (2013).
- [3] R. Soomro, M.J. Ahmed M. Memon, *Turkish Journal of Chemistry*. 35, 155 (2011).
- [4] R.V. Lebedeva, A.A. Leont'eva, A.N. Tumanova, N.I. Mashin, *Inorganic Materials*. 46, 323 (2010).
- [5] E.K. Paleologos, C.D. Stalika, *Analyst*. 126, 389(2001).
- [6] K. Mortazavi, M. Ghaedi, M. Montazerzohori, and M. Roosta, *Fresenius Environ. Bull*, 20, 2847 (2011).
- [7] M. Ghaedi, K. Mortazavi, M. Jamshidi, M. Roosta, and B. Karami, *Toxicol. Environ. Chem*, 94, 846 (2012).
- [8] H. Ebrahimzadeh, A.A. Asgharinezhad, N. Tavassoli, O. Sadeghi, M.M. Amini, F. Kamarei, *International Journal of Environmental Analytical Chemistry*. 92, 509(2012).
- [9] M.E. Mahmoud, ME, Yakout AA, Ahmed SB and Osman MM. *azardous Materials*. 158, 541(2008).
- [10] World Health Organization, Trace Elements in Human Nutrition and Health, Geneva, 163,167(1996).
- [11] Z. Marczenko, Separation and Spectrophotometric Determination of Elements, Horwood, Chichester,(1986).
- [12] B.Chiswell, G.Rauchle, M.Pascoe; *Talanta*, 37, 237 (1990).
- [13] J.Liu, Y.Feng, G.Jiang; *JAOAC Int*. 84, 1179 (2001).
- [14] Q.Wei, L.Yan, G.Chang, Q.Ou; *Talanta*, 59, 253 (2003).
- [15] I.F.Dolmanova, G.A.Zolotova, M.A.Ratina; *J. Anal. Chem.*, USSR, 33, 1063 (1978).
- [16] L.Jianli, W.Budong, Z.Fenyan; *Analyst*, 118, 1213 (1993).
- [17] R.Liu, A.Zhang, D.Liu, S.Wang; *Analyst*, 120, 1195 (1995).
- [18] K.Watanabe, S.Takahashi, M.Itagaki; *Anal.Chim. Acta*, 522, 281 (1996).
- [19] L.Su, J.Li, H.Ma, G.Tao; *Anal.Chim.Acta*, 522, 281 (2004).
- [20] J.RuzickaE.H.Hansen; *Flow Injection Analysis*, 2nd ed., Wiley, New York. (1988).
- [21] M.S. Hosseini, and R. Hassan-Abadi, *Anal Chim*, 97, 1269 (2007).
- [22] O. Dalman, V.N. Bulut, I. Degirmencioglu, and M. Tufekci, *Turk J Chem*, 31, 631 (2007).
- [23] J. Yang, C. Ma, S. Zhang, Z. Shen; *Anal.Chim.Acta*, 235, 323 (1990).
- [24] C. Zhang, S. Kawakubo, T. Fukasawa; *Anal.Chim. Acta*, 217, 23 (1989).
- [25] S. Nakano, M. Nozawa, M. Yanagawa; *Anal.Chim. Acta*, 261, 183 (1992).
- [26] T. Yamane, K. Koshino; *Talanta*, 43, 963 (1996).
- [27] S. Nakano, K. Tanaka, R. Oki, T. Kawashima, *Talanta*, 49, 1077 (1999).
- [28] N. Maniasso, E. A. G. Zaggatto; *Anal.Chim.Acta*, 366, 87 (1998).
- [29] (a): F. Ahmadi, E. Niknam, K. Niknam, and A. Khanmohammadi, *Arabian Journal for Science and Engineering*, 36, 47 (2011), (b): A. Shokrollahi, H.E. Haghghi, E. Niknam, and K. Niknam, *Quím. Nova São Paul*, 36, 3, 273 (2013).
- [30] A. Shokrollahi, M. Ghaedi, H. Ghaedi, *J. Chin. Chem. Soc*. 54, 933(2007).
- [31] M. Noroozifar, M.K. Motlagh, *Analytical Sciences*. 19, 705(2003).
- [32] P. Venkateswaran, K. Palanivelu, *Separation and Purification Technology*, 40, 279(2004).
- [33] Y. Akama, A. Sali, *Talanta*. 57, 681(2002).

- [34] IUPAC, Nomenclature, *Pure Appl. Chem.* 45, 105(1976).  
[35] F.Ahmadi, G. Haghdoost, G. Shafiee, A. Beydokhti, Trade Science Inc. – INDIA 36, 321 (2008).  
[36] B. Mandal, U.S. Roy, *Indian J. Chem. A* 47, 1497(2008).  
[37] M. Ghaedi, E. Asadpour, A. Vafaie, *J. Molecular and Biomolecular Spectroscopy.*63, 182(2006).  
[38] B.W. Yang, Z.F. Fan, *Atom. Spectrosc.*29, 193(2008).  
[39] D.H. Chen, M. He, C.Z. Huang, B. Hu, *Atom.Spectrosc.*29, 165(2008).