



Scholars Research Library

Der Pharma Chemica, 2012, 4(5):1902-1905
(<http://derpharmachemica.com/archive.html>)



ISSN 0975-413X
CODEN (USA): PCHHAX

Study on facile method for cellulose degradation from waste paper

Prasanna S Pande

Department of Chemistry, Shankarlal Khandelwal College, Godbole Plots, Akola (M.S.)

ABSTRACT

The facile method the hydrolysis of cellulose from waste paper material has been worked out. Very mild conditions are used for cellulose breakdown to glucose. The glucose produced in this technique was also estimated qualitatively as well as quantitatively. The production of ethanol from the glucose was also studied qualitatively.

Keywords: Cellulose degradation, Glucose

INTRODUCTION

Literature survey reveals that The basic material for the production of first generation biofuels are often seeds or grains such as wheat, which yields starch that is fermented into bioethanol. There are two common strategies of producing biofuels. One is to grow crops high in sugar like sugar cane, sugar beet, and sweet sorghum and then use yeast fermentation to produce ethyl alcohol. The second is to grow plants that contain high amounts of vegetable oil, such as soybean, jatropha, or pongamia pinnata. When these oils are heated, their viscosity is reduced, and they can be burned directly in a diesel engine, or they can be chemically processed to produce fuels such as biodiesel. Wood and its by products can also be converted into biofuels such as methanol or ethanol fuel [1].

Second-generation biofuel production processes use a variety of non food crops. These include waste biomass, the stalks of wheat, corn, wood, etc. Second generation biofuels use biomass including cellulosic biofuels from non food crops [2]. Many second generation biofuels are under development such as biohydrogen, biomethanol, DMF, biohydrogen diesel, mixed alcohols and wood diesel. Cellulosic ethanol production uses non food crops or inedible waste products and does not divert food away from the animal or human food chain.. This feedstock is abundant and diverse, and in some cases it is a significant disposal problem.

The work on biofuels like butanol and ethanol has tremendous international recognition. There are international organizations such as IEA Bioenergy, established in 1978 by the OECD International Energy Agency (IEA), with the aim of improving cooperation and information exchange between countries that have national programs in bioenergy research, development and deployment. The U.N. International Biofuels Forum is formed by Brazil, China, India, South Africa, the United States and the European Commission[3]. A recent publication by the European Union highlighted the potential for waste-derived bioenergy to contribute to the reduction of global warming. The report concluded that the equivalent of 19 million tons of oil is available from biomass by 2020, 46% from bio-wastes: municipal solid waste (MSW), agricultural residues, farm waste and other biodegradable waste streams [4,5].

It is worth experimenting to devise a facile methodology for the degradation of cellulose from waste paper and to find a way ethanol production.

MATERIALS AND METHODS

Waste paper was collected from the campus of Shankarlal Khandelwal college Akola.

Pre-treatment method and estimation of glucose

01 gram of waste paper was treated with 50 ml of 10 % HCl solution in a round bottom flask and heated for 1 hour at 85°C the reaction was stopped after 1 hour. The reaction mixture was allowed to cool and filtered. The in the sample was neutralized by small volume of concentrated potassium hydroxide solution. The in situ production of glucose was estimated by felhing solution method as below

Chemicals used

0.06 N standard glucose solution, felhing solution A, felhing solution B

Standardization of felhing solution

50 ml of felhing solution A and 50 ml of felhing solution B was mixed in a 250ml beaker out of which 10 ml of the solution was taken in a clean china dish with the help of pipette. The china dish was kept on heating mental and the solution was heated to boiling. To this felhing solution, the standard glucose solution of normality 0.06N was added drop wise through the burette till the colour changes from dark blue to brick red.

Observations

Sr. No.	Volume of Felhing solution	Volume of Std. Glucose	End point
1	10ml	5.3ml	
2	10ml	5.4ml	5.4ml
3	10ml	5.4ml	

Calculation

Normality of felhing solution

Glucose Felhing

$$N_1V_1 = N_2V_2$$

$$0.06 \times 5.4 = N_2 \times 10$$

$$N_2 = \frac{0.06 \times 5.4}{10}$$

$$N_2 = 0.0324 \text{ N}$$

For experimental solution

Sr. No.	Volume of Felhing solution	Volume of Experimental Glucose	End point
1	10ml	12.3ml	
2	10ml	12.4ml	12.4ml
3	10ml	12.4ml	

To calculate normality of experimental glucose solution

Felhing Experimental glucose

$$N_2V_2 = N_3V_3$$

$$0.032 \times 10 = N_3 \times 12.4$$

$$N_3 = \frac{0.032 \times 10}{12.4}$$

$$N_3 = 0.025 \text{ N}$$

Normality of experimental glucose = 0.025 N

Wt/lit = Normality x eq. Wt

$$\text{Wt/lit} = 0.025 \times 90$$

$$\text{Wt/lit} = 2.25 \text{ g}$$

Per 50 ml

$$50 \times 2.25$$

$$1000$$

$$= 0.11 \text{ g}$$

Result :- 0.11 g glucose (11%)

Scaling up experiment

10 gram of waste paper was treated with 500 ml of 10 % HCl solution in a round bottom flask and heated for 1 hour at 85°C the reaction was stopped after 1 hour. The reaction mixture was allowed to cool and filtered. The in the sample was neutralized by small volume of concentrated potassium hydroxide solution. The in situ production of glucose was estimated by felhing solution method as below

Chemicals used

0.06 N standard glucose solution, felhing solution A, felhing solution B

Standardization of felhing solution

50 ml of felhing solution A and 50 ml of felhing solution B was mixed in a 250ml beaker out of which 10 ml of the solution was taken in a clean china dish with the help of pipette. The china dish was kept on heating mental and the solution was heated to boiling. To this felhing solution, the standard glucose solution of normality 0.06N was added drop wise through the burette till the colour changes from dark blue to brick red.

Observations

Sr. No.	Volume of Felhing solution	Volume of Std. Glucose	End point
1	10ml	5.3ml	
2	10ml	5.4ml	5.4ml
3	10ml	5.4ml	

Calculation

To calculate normality of felhing solution

Glucose Felhing

$$N_1V_1 = N_2V_2$$

$$0.06 \times 5.4 = N_2 \times 10$$

$$N_2 = \frac{0.06 \times 5.4}{10}$$

$$N_2 = 0.0324 \text{ N}$$

For experimental solution

Sr. No.	Volume of Felhing solution	Volume of Experimental Glucose	End point
1	10ml	13.3ml	
2	10ml	13.4ml	13.4ml
3	10ml	13.4ml	

To calculate normality of experimental glucose solution

Felhing Experimental glucose

$$N_2V_2 = N_3V_3$$

$$0.032 \times 10 = N_3 \times 13.4$$

$$N_3 = \frac{0.032 \times 10}{13.4}$$

$$N_3 = 0.023 \text{ N}$$

Normality of experimental glucose = 0.023 N

Wt/lit = Normality x eq. Wt

$$\text{Wt/lit} = 0.023 \times 90$$

$$\text{Wt/lit} = 2.07 \text{ g}$$

Per 500 ml

$$500 \times 2.07$$

$$\frac{1000}{1000}$$

$$= 1.035 \text{ g}$$

Result :- 1.035 g glucose (i.e 10.35%)

Production of ethanol by fermentation process

The solution obtained from experiment no. 04 was filtered and concentrated up to 100 ml. This solution was labeled as Sample A. Basal growth medium was prepared as below.

Yeast extract 4.5 g, bromothymol blue 30 g, peptone 7.5 g, distilled water 1000 ml, Chloramphenicol 70 mg (added after autoclaving),

A tube containing 10 ml of basal medium was prepared and autoclaved. 5 ml of sample A was added to the tube under aseptic condition. The tube was then inoculated with active culture of yeast. The mixture was then incubated at 28^oC for 12 days. After 12 days, the contents were filtered and 10 ml of the solution was taken in a test tube. To this solution 5 drops of 10% solution NaOH solution was added. With a separate dropper, KI solution was added drop wise till the solution became yellow. The mixture was then allowed to stand for 5 minutes and the pulsated vigorously. Yellow precipitate was obtained which is the indication of ethyl alcohol.

Acknowledgements

The author is thankful to the Management and the Principal of Shankarlal Khandelwal College, Akola for providing the necessary facilities and encouragement. The author is thankful to the UGC (WRO), Ganeshkhind, Pune for providing the financial assistance for the minor research project on this research problem.

REFERENCES

- [1] Pu, Yunqiao; Dongcheng Zhang, Preet M. Singh and Arthur J. Ragauskas (2007). *Biofuels, Bioproducts and Biorefining (BioFPR)* 2 (1): 58–73.
- [2] Chris Somerville. "Development of Cellulosic Biofuels". U.S. Dept. of Agriculture.
<http://www.usda.gov/oce/forum/2007>
- [3] "Press Conference Launching International Biofuels Forum". United Nations Department of Public Information. 2007.
http://www.un.org/News/briefings/docs/2007/070302_Biofuels.doc.htm.
- [4] European Environment Agency (2006) How much bioenergy can Europe produce without harming the environment?
- [5] Marshall, A. T. (2007) Bioenergy from Waste: A Growing Source of Power, *Waste Management World Magazine*, April, p34-37 .