

ISSN 0975-413X CODEN (USA): PCHHAX

Der Pharma Chemica, 2018, 10(8): 96-101 (http://www.derpharmachemica.com/archive.html)

Synthesis and Biological Activities of [1,3]-Oxazine Derivatives

Chaitra G^{*}, Rohini RM

Department of Pharmaceutical Chemistry, Al-Ameen College of Pharmacy, Bengaluru, India

ABSTRACT

A series of novel [1,3]-oxazine derivative of N-[4-(2-Amino-4-phenyl-6H-[1,3]oxazine-6-yl)-phenyl]-nicotinamide are synthesized due to its wide range of biological activities. To achieve the target compounds the starting material used was nicotinyl chloride. Chalcone derivatives were synthesized by carrying the reaction of benzaldehyde derivatives with p-amino acetophenone. The obtained products were allowed to react with urea to give heterocyclic derivatives of oxazine 5(a-f) respectively. The structures of all synthesized molecules were confirmed by Ultraviolet (UV), Infrared (IR), Proton Nuclear Magnetic Resonance (¹H-NMR), Carbon-13 Nuclear Magnetic Resonance (¹³C-NMR) and Mass spectral data. They were screened for their in-vitro anti-inflammatory activity by bovine serum albumin and protease method and anti-oxidant activity by diphenyl picryl hydrazide and nitric oxide method. N-{4-[2-Amino-4-(3,4,5-trimethoxy-phenyl)-6H-[1,3]oxazine-6-yl]-phenyl}-nicotinamide 5(c) and N-{4-[2-Amino-4-(3-nitro-phenyl)-6H-[1,3]oxazine-6-yl]-phenyl}-nicotinamide for BSA and protease methods for anti-inflammatory activity; DPPH and NO methods for anti-oxidant method at 10 µg, 50 µg and 100 µg.

Keywords: Oxazine, Chalcone, Antiinflammatory, Antioxidant activity.

INTRODUCTION

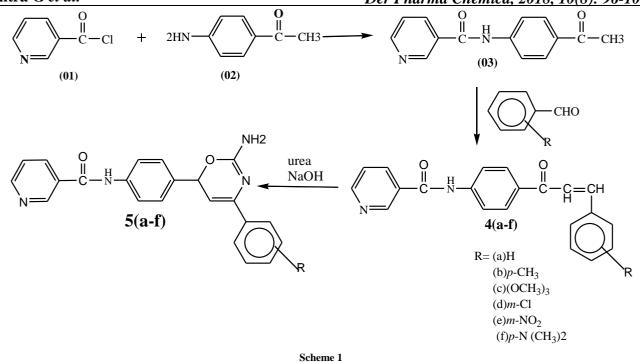
Heterocyclic derivatives of chalcones that have been synthesized have shown biological and pharmacological activities, resulting good chemotherapeutics compounds [1]. Oxazine derivatives are one among the heterocyclic compounds that shows biological activities [2] like anti-hyperglycemic [3], anti-leishmanial [4], anti- tubercular [5], anti-ulcer [6], anti-cancer [7]. Oxazine are heterocyclic compounds containing one nitrogen and one oxygen with three isomeric forms [8]. Anti-oxidant and anti-inflammatory based drug are used for the prevention and treatment of complex diseases like atherosclerosis, stroke, diabetes, alzheimer's disease and cancer [9]. The interest on 1,3-oxazine molecules has increased recently because compounds containing dihydro-1,3-oxazine ring system exhibited a wide spectrum of pharmacological activities such as anti-malarial [10], anti-tumor [11], anti-bacterial [12-14], anti-oxidant [15-17], anti-inflammatory activity [18] and their versatility as synthetic intermediates [19]. Particular attention has been paid to these compounds since the discovery of the non-nucleoside reverse transcriptase inhibitor trifluoromethyl-1,3 -oxazine-2-one, which shows high activity against a variety of HIV-1 mutant strains [20]. This has been the prime step for the synthesis of various compounds consolidating the 1,3 -oxazine moiety [21].

MATERIALS AND METHODS

Melting point of the synthesized compounds was determined in electro thermal apparatus using fused capillary tubes. Monitoring of the reaction and the purity of the compounds was checked by thin-layer chromatography using silica gel G plates of 0.5 mm thickness as stationary phase in combination of *n*-hexane: ethyl acetate in different ratios as mobile phase. The spots were visualized by using iodine chamber and Ultraviolet (UV) chamber (Scheme 1).

The UV spectra of the synthesized compounds were recorded on Shimadzu UV-1601 and the values of wavelength (λ_{max}) were reported in nm. The Infrared (IR) spectra of the synthesized compounds were recorded on a Fourier Transform IR spectrophotometer (model Shimadzu 8700) in the range of 4000-400 cm⁻¹ using KBr pellets and value of λ_{max} are reported in cm⁻¹ and the spectra were interpreted. Proton Nuclear Magnetic Resonance (¹H-NMR) and Carbon-13 Nuclear Magnetic Resonance (¹³C-NMR) spectra were recorded on Bruker Avance II 400 NMR spectrometer using CDCl₃. Chemical shift (δ) are reported in parts per million downfield from internal reference, Tetramethylsilane (TMS) and the spectra were interpreted. Mass spectra were recorded on Mass spectrophotometer (model Shimadzu) by MS-2010A. Chaitra G et al.

Der Pharma Chemica, 2018, 10(8): 96-101



Preparation of N-(4-Acetyl-phenyl)-nicotinamide (03)

To an ice cold solution of nicotinyl chloride (0.01 mol) dissolved in methanol, a methanolic solution of *p*-aminoacetophenone (0.01 mol) was added dropwise with constant stirring. Stirring was continued for 15 min then refluxed for one hour. The reaction mixture was cooled and the obtained precipitate was filtered. Crude product was recrystallized from isopropyl alcohol.

General method for synthesis of substituted 4 (a-f)

Equimolar quantity of *N*-(4-Acetylphenyl)-nicotinamide (0.01 mol) and substituted benzaldehyde (0.01 mol) were dissolved in absolute alcohol and 40% KOH solution was added slowly with stirring. Continuing the reaction for 9 h and left overnight. The reaction mixture was decomposed in ice-water to obtain product. Crude product was recrystallized from ethanol.

General method for the synthesis of substituted 5 (a-f)

Equimolar quantity of 4 (a-f) and urea were dissolved in ethanolic NaOH and was stirred for 2-3 h at room temperature. Refluxed for 6 h and poured into cold water with continuous stirring for 1 h and was cooled at 0°C for 42 h. The precipitate obtained was filtered, washed and recrystalized from ethanol.

N-(4-Acetyl-phenyl)-nicotinamide (3)

Orange color; M. p. 128°C; Yield: 64.5%; IR (KBr) cm⁻¹: (C=O) 1736.58 cm⁻¹, (Ar C=C) 1427.07 cm⁻¹, (Ar C-H) 3175.22 cm⁻¹, (C=N) 1677.77 cm⁻¹, (CH3) 2792.42 cm⁻¹, (CONH) 1575 cm⁻¹; ¹H-NMR (CDCl₃-ppm): δ (ppm)=8.0 (d, 1H, CONH), δ (ppm)=8.84 (m, 4H, Ar), δ (ppm)=7.75 (d, 2H, Ar), δ (ppm)=7.84 (d, 2H, Ar), δ (ppm)=2.55 (s, 3H, CH₃).

N-[4-(3-Phenyl-acryloyl)-phenyl]-nicotinamide 4(a)

Pale yellow color; M. p. 121°C; Yield: 69.28%; IR (KBr) cm⁻¹: (NH) 3359 cm⁻¹, (Ar C-H) 3025.76 and 3083.62 cm⁻¹, (CH₃)2917.77 cm⁻¹, (CONH) 1515 cm⁻¹, (C=C) 1643 cm⁻¹, (Ar C=C) 1448.28 cm⁻¹, (C=O) 1604.48 cm⁻¹.

N-[4-(2-Amino-4-phenyl-6H-[1,3]oxazine-6-yl)-phenyl]-nicotinamide 5(a)

Pale yellow color; M. p. 106°C; Yield: 63.6%; λ_{max} : 379.00 nm; IR (KBr) cm⁻¹: (NH₂) 3468 cm⁻¹, (NH) 3355.53 cm⁻¹, (Ar C-H) 3027.69 cm⁻¹, (C-O-C) 1035.59 cm⁻¹, (C=N) 1646 cm⁻¹, (Ar C=C) 1596 and 1492 cm⁻¹, (CONH) 1529 cm⁻¹; 1H-NMR (CDCl₃-ppm): δ (ppm)=7.96 (d, 1H, CONH), δ (ppm)=7.57 (d, 2H, ArH), δ (ppm)=7.17 (m, 1H, ArH), δ (ppm)=7.30 (d, 2H, ArH), δ (ppm)=7.21 (s, 2H, ArH), δ (ppm)=2.0 (s, 2H, NH₂), δ (ppm)=6.7 (d, 1H, C=C), δ (ppm)=5.19 (d, 1H, CH oxazine), δ (ppm)=5.19 (d, 1H, C-O), δ (ppm)=7.96 (m, 4H, 4-pyridine), δ (ppm)=7.17 (m, 4H, 1-benzene) δ (ppm)=2.0 (d, 2H, NH₂).

N-[4-(2-Amino-4-p-tolyl-6H-[1,3]oxazine-6-yl)-nicotinamide 5(b)

Pale yellow color; M. p. 89°C; Yield: 92.33; λ_{max} : 326.50; IR (KBr) cm⁻¹: (NH₂) 3462.56 cm⁻¹, (NH) 3343.96 cm⁻¹, (Ar C-H) 3046.98 cm⁻¹, (C-O-C) 1075 cm⁻¹, (C=N) 1600.63 cm⁻¹, (Ar C=C) 1611 and 1438.64 cm⁻¹, (CONH) 1511.92 cm⁻¹, (CH₃) 2918.73 cm⁻¹; ¹H-NMR (CDCl₃-ppm): δ (ppm)=7.93 (d, 1H, CONH), δ (ppm)=7.51 (d, 2H, ArH), δ (ppm)=7.2 (m, 1H, ArH), δ (ppm)=7.4 (d, 2H, ArH), δ (ppm)=7.29 (s, 2H, ArH), δ (ppm)=2.40 (s, 2H, NH₂), δ (ppm)=3.73 (d, 3H, CH₃), δ (ppm)=6.69 (d, 1H, C=C), δ (ppm)=5.2 (d, 1H, CH oxazine), δ (ppm)=6.68 (d, 3H, C-O).

N-{4-[2-Amino-4-(3,4,5-trimethoxy-phenyl)-6H-[1,3]oxazin-6-yl]-phenyl}-nicotinamide 5(c)

Yellow color; M. p. 156°C; Yield: 98.2; λ_{max} : 325.60 nm; IR (KBr) cm⁻¹: (NH₂) 3457.74 cm⁻¹, (NH) 3365.17 cm⁻¹, (Ar C-H) 3232.1 cm⁻¹, (C-O-C) 1126.22 cm⁻¹, (C=N) 1628.59 cm⁻¹, (Ar C=C) 1611 and 1438 cm⁻¹, CONH 1593.88 cm⁻¹, OCH₃ 2997.8, 2938.98, 2832.92 cm⁻¹; ¹H-NMR (CDCl₃-ppm): δ (ppm)=7.9 [1H, (d, Ar H)], δ (ppm)=7.6 [1H, (s, Ar H)], δ 7.7 [1H (d, Ar H)], δ (ppm)=7.7 [1H, (d, Ar H)], δ (ppm)=7.4 [2H, (s, -C=C)], δ (ppm)=7.2 [2H, (s, -C=C)], δ (ppm)=6.1 [1H, (s, -C-H)], δ (ppm)=6.7 [1H, (d, -C=C)], δ (ppm)=2.41 [2H, (s, -NH₂)], δ (ppm)=6.4 [2H, (d, Ar H)], δ (ppm)=3.7 [9H, (m, -OCH₃)]; ¹³C-NMR (CDCl₃-ppm): (2C, aromatic)-152.97, (4C, oxazine, C=N) δ (ppm)=151.61, 151.54,

Chaitra G et al.

Der Pharma Chemica, 2018, 10(8): 96-101

(1C, oxazine, C=O) δ (ppm)=197.17, (1C, oxazine, C-NH₂) δ (ppm)=188.16, (1C,C=C) δ (ppm)=121.34, (1C,C-C) δ (ppm)=128.04, (1C, ArH) δ (ppm)=137.05, (2C, ArH) δ (ppm)=127.41 and 127.18, (2C, ArH) δ (ppm)=121.34, (1C, C-N) δ (ppm)=137.05, (1C, C-O) δ (ppm)=64.36, (1C,C-N) δ (ppm)=130.85, (2C, ArH) δ (ppm)=105.44, (3C, OCH₃) δ (ppm)=56.18, 56.09 and 56.01; m/e: 476.0 (molecular ion).

N-{4-[2-Amino-4-(3-chloro-phenyl)-6H-[1,3]oxazin-6-yl]-phenyl}-nicotinamide 5(d)

Yellowish; M. p.179°C; Yield: 70.0; λ_{max} : 370.40 nm; IR (KBr) cm⁻¹: (NH₂) 3419 cm⁻¹, (NH) 3332 cm⁻¹, (Ar C-H) 3052 cm⁻¹, (C-O-C) 1126.29 cm⁻¹, (C=N) 1634 cm⁻¹, (Ar C=C) 1442.49 cm⁻¹, (CONH) 1518.3 cm⁻¹; ¹H-NMR (CDCl₃-ppm): δ (ppm)=7.81 (d, 1H, CONH), δ (ppm)=7.52 (d, 2H, ArH), δ (ppm)=7.2 (m, 1H, ArH), δ (ppm)=7.4 (d, 2H, ArH), δ (ppm)=7.3 (s, 2H, ArH), δ (ppm)=2.0 (s, 2H, NH₂), δ (ppm)=6.69 (d, 1H, C=C), δ (ppm)=5.1 (d, 1H, CH oxazine), δ (ppm)=6.68 (d, 3H, C-O).

N-{4-[2-Amino-4-(3-nitro-phenyl)-6H-[1,3]oxazin-6-yl]-phenyl}-nicotinamide methane 5(e)

Dark yellow; M. p. 159°C; yield: 72.4; λ_{max} : 374.60 nm; IR (KBr)cm⁻¹: (NH₂) 3425.92 cm⁻¹, (NH) 3338.18 cm⁻¹, (Ar C-H) 3089 cm⁻¹, (C-O-C) 1176.36 cm⁻¹, (C=N) 1634.38 cm⁻¹, (Ar C=C) 1444.42 cm⁻¹, (CONH) 1532.17 cm⁻¹; ¹H-NMR (CDCl₃-ppm): δ (ppm)=7.83 (d, 1H, CONH), δ (ppm)=7.59 (d, 2H, ArH), δ (ppm)=7.31 (m, 1H, ArH), δ (ppm)=7.54 (d, 2H, ArH), δ (ppm)=7.29 (s, 2H, ArH), δ (ppm)=2.20 (s, 2H, NH₂), δ (ppm)=6.69 (d, 1H, C=C), δ (ppm)=5.2 (d, 1H, CH oxazine), δ (ppm)=6.68 (d, 3H, C-O).

N-{4-[2-Amino-4-(3-dimethylamino-phenyl)-6H-[1,3]oxazin-6-yl]-phenyl}-nicotinamide methane 5(f)

Dark yellow; M. p. 190°C; yield: 72.5; λ_{max} : 412.60 nm; IR (KBr) cm⁻¹: (NH₂) 3400.85 cm⁻¹, (NH) 3334.32 cm⁻¹, C-H (Ar) 3046.98 cm⁻¹, C-O-C 1028.84 cm⁻¹, C=N 1598.7 cm⁻¹, C=C (Ar) 1432.85 cm⁻¹, (CONH) 1524.45 cm⁻¹, [(CH₃)₃] 2807.85, 2911.99 cm⁻¹; ¹H-NMR (CDCl₃-ppm): δ (ppm)=7.90 (d, 1H, CONH), δ (ppm)=7.50 (d, 2H, ArH), δ (ppm)=7.29 (m, 1H, ArH), δ (ppm)=7.4 (d, 2H, ArH), δ (ppm)=7.32 (s, 2H, ArH), δ (ppm)=2.0 (s, 2H, NH₂), δ (ppm)=6.69 (d, 1H, C=C), δ (ppm)=5.19 (d, 1H, CH oxazine), δ (ppm)=6.68 (d, 3H, C-O), δ (ppm)=2.85 [m, 6H, N(CH₃)₂].

In vitro anti-inflammatory activity

Bovine serum albumin (BSA) method [22]

A solution of 0.2% w/v of Bovine serum albumin BSA was prepared in Tris buffer saline. Test sample stock solutions of 1000 μ g/ml was prepared. From stock solutions two different concentrations of 100 μ g/ml and 200 μ g/ml were prepared by using methanol as a solvent. To 0.1 ml of each test sample 5 ml of 0.2% BSA was added. The blank consisted 5 ml 0.2% w/v BSA solution and 0.1 ml methanol. The positive control consist 0.1 ml (100 μ g/ml) of Indomethacin in methanol and 5 ml 0.2% w/v BSA solution. The test samples were heated at 72°C for five minutes and then cooled for 10 min. The absorbance of these solutions was determined by using spectrophotometer at a wavelength of 660 nm. The denaturation of the protein was determined on a percentage basis relative to the blank control using the following formula:-

% inhibition = $\frac{\text{Absorbance of Control} - \text{Absorbance of Test}}{\text{Absorbance of Control}} \times 100$

Protease inhibition assay method [23]

The reaction mixture containing 0.5 ml of protease solution (0.06 mg of trypsin in 20 mM of Tries HCl buffer), 0.5 ml of Tris HCl buffer (20 mM) and 1 ml of test sample of different concentration was prepared. The mixture was incubated at 37° C for 15 min and then 1 ml of 0.8% w/v casein was added. The mixture was incubated for an additional 20 min. Then 2 ml of 70% perchloric acid was added to terminate the reaction. Cloudy suspension was centrifuged and the absorbance of the supernatant was read at 210 nm against buffer as blank. The % inhibition was calculated using following formula:-

% inhibition =
$$\frac{\text{Absorbance of Control} - \text{Absorbance of Test}}{\text{Absorbance of Control}} \times 100$$

In vitro antioxidant activity

Diphenyl picryl hydrazide (DPPH) method [24]

A stock solution of DPPH (10 mg) was prepared by dissolving in 10 ml of methanol. From this stock solution, different dilutions were made to obtain concentrations of 10 to 100 μ g/ml. The absorbance was recorded at 517 nm.

Standard solution ascorbic acid (10 mg) was dissolved in 10 ml of methanol. From this stock solution dilutions 10 to 100 μ g/ml were measured. To ml of different concentration of ascorbic acid 1 ml of DPPH solution of 30 μ g/ml concentration was added and volume was made up to 10 ml with methanol. The absorbance was recorded for these dilutions at 517 nm after incubation for 30 min. The effect of ascorbic acid (vitamin C) on DPPH was assessed for comparison with that of synthesized compounds.

The test samples were prepared by initial preparation of stock solution of $(1000 \ \mu g/ml)$ in methanol and different concentrations were prepared (100, 50 and 10 $\mu g/ml$). To all these dilutions, 1 ml of DPPH solution (30 $\mu g/ml$) was added and absorbance were recorded at 517 nm after duration of 30 min. Percentage inhibition of free radical activity was calculated using the following formula.

% inhibition = $\frac{\text{Absorbance of Control} - \text{Absorbance of sample}}{\text{Absorbance of Control}} \times 100$

Nitric oxide radical scavenging method [25]

To 0.5 ml of 10 mM sodium nitroprusside (10 mM) in phosphate buffer saline was added to 1 ml of test sample of different concentration, incubated at 25°C for 180 min then added 1 ml of griess reagent [Solution A: 1% sulphanilamide in 2.5% phosphoric acid; Solution B: 0.1% napthyl ethylene diamine in 2.5% phosphoric acid].

Solution A and solution B were mixed in equal volume and were used before 12 h. The positive control ascorbic acid solution was prepared similarly and was incubated. A blank solution was prepared. The absorbance of chromophore formed was measured at 546 nm on UV-Visible spectroscopy.

NO Scavenging $(\%) = \frac{\text{Absorbance of Control} - \text{Absorbance of sample}}{\text{Absorbance of Control}} \times 100$

RESULTS AND DISSCUSSION

Anti-inflammatory activity

Bovine serum albumin (BSA) method

Among the screened compounds N-{4-[2-Amino-4-(3,4,5-trimethoxy-phenyl)-6*H*-[1,3]oxazine-6-yl]-phenyl}-nicotinamide 5(c) and N-{4-[2-Amino-4-(3-nitro-phenyl)-6*H*-[1,3]oxazine-6-yl]-phenyl}-nicotinamide methane 5(e) showed good activity compared to positive control of indomethacin. Other compounds showed dose dependent good *in vitro* anti-inflammatory activity as compared to standard indomethacin at 200 μ g/ml. the results have been depicted in Table 1.

Table 1: Anti-inflammatory activity by BSA method

| Commonweak | Percentage Inhibition (%) | | |
|---------------|---------------------------|------------------|--|
| Compound code | Dose (100 µg/ml) | Dose (200 µg/ml) | |
| Indomethacin | 78.01 | 94.8 | |
| 5a | 53.3 | 73 | |
| 5b | 53.3 | 66.6 | |
| 5c | 66.6 | 86.6 | |
| 5d | 46.6 | 86.6 | |
| 5e | 73.3 | 86.6 | |
| 5f | 66.6 | 80.6 | |

Proteases method

All the synthesized compounds were screened for *in-vitro* anti-inflammatory activity by proteases method. It was evident by the observation that the compound 5(c) and 5(e) exhibited excellent anti-inflammatory activity. The results have shown in following Table 2.

| Table 2: Anti-inflammatory activi | ty by Protease method |
|-----------------------------------|-----------------------|
|-----------------------------------|-----------------------|

| Compound code | Percentage Inhibition (%) | | |
|---------------|---------------------------|-----------------|-----------------|
| | Dose (100 µg/ml) | Dose (50 µg/ml) | Dose (10 µg/ml) |
| Indomethacin | 89.32 | 69.31 | 30.15 |
| 5a | 77.6 | 70.4 | 52.22 |
| 5b | 77.88 | 57.64 | 50.55 |
| 5c | 77.74 | 71.43 | 67.64 |
| 5d | 77.6 | 66.33 | 64.89 |
| 5e | 86.34 | 68.91 | 68.77 |
| 5f | 54.74 | 71.01 | 61.66 |

Anti-oxidant activity

Diphenyl picryl hydrazide (DPPH) method

The hydrogen atom donating ability of the synthesized was determined by the decolorization of methanol solution of 2,2-diphenyl-1picrylhydrazyl (DPPH). DPPH produced violet color in methanol and faded to yellow color in the presence of antioxidants. Compared to standard ascorbic acid 5(c) and 5(e) compounds exhibited good activity. The results have shown in below Table 3.

Table 3: Anti-oxidant activity by DPPH method

| Compound code | Percentage Inhibition (%) | | |
|---------------|---------------------------|-----------------|-----------------|
| | Dose (100 µg/ml) | Dose (50 µg/ml) | Dose (10 µg/ml) |
| Ascorbic acid | 99.25 | 95.41 | 90.5 |
| 5a | 94.1 | 90.6 | 89.4 |
| 5b | 98.6 | 86 | 85.9 |
| 5c | 94.7 | 94.4 | 94 |
| 5d | 87 | 82.6 | 80 |
| 5e | 90 | 85 | 84.6 |
| 5f | 76.6 | 62.2 | 61.5 |

Nitric oxide (NO) method

The compounds have been tested in 10, 50 and 100 μ g/ml which have shown good activity among them 5(c) and 5(e) exhibited highest activity as compared to standard ascorbic acid. The results have depicted in Table 4.

| Compound code | Percentage Inhibition (%) | | |
|---------------|---------------------------|-----------------|-----------------|
| | Dose (100 µg/ml) | Dose (50 µg/ml) | Dose (10 µg/ml) |
| Ascorbic acid | 81.112 | 80.4 | 78.81 |
| 5a | 81.11 | 81.11 | 81.73 |
| 5b | 80.48 | 81.11 | 81.11 |
| 5c | 81.11 | 81.73 | 81.11 |
| 5d | 89.11 | 85.11 | 81.11 |
| 5e | 81.11 | 81.11 | 81.11 |
| 5f | 81.11 | 83.46 | 81.11 |

Table 4: Anti-oxidant activity by NO method

CONCLUSION

From our present study, it can be concluded that six derivatives of substituted 1,3-oxazine were obtained from the intermediate chalcone on treatment with urea in presence of ethanolic NaOH. The compounds were synthesized in good percentage of yield their physical and analytical determination was done by using melting point apparatus, purification of compounds by TLC. The structural assignments of these compounds were identified and characterized by using IR, ¹H-NMR, ¹³C-NMR and Mass techniques. It is evident from the research work that the series of synthesized and screened compounds can further be explored for *in vivo* studies.

REFERENCES

- [1] P. Anusha, P. Mani Chandrika, S. Shruthi, S.U. Nishat, S. Sultana, World. J. Pharm. Pharm. Sci., 2015, 4(11), 885-895.
- [2] G.R. Kamala, L. Patnaik, M. Annapurna, S. Sasanka, M.S.K. Naidu, J. Med. Chem. Drug. Discovery., 2016, 8, 2332.
- [3] H. Jamal, W.H. Ansari, S.J. Rizvi, Biol. Med., 2009, 1, 107-115.

[4] Y. Zuo, Euro. J. Med. Chem., 2012, 50, 393-404.

- [5] V. Kotra, India. J. chem., 2010, 49, 1109-1116.
- [6] N.S. Pamar, India. J. Phy. Pharma. Col., 1998, 42, 343-351.
- [7] H.P. Singh, Sch. Res. Lib., 2010, 2, 460-472.
- [8] F.V. Lauro, D.C. Franciso, L.R. Marcela, G.C. Elodia, P.G. Eduardo, LR. Maria, J. Chem., 2014, 1-9.

[9] K. Elumalai, M.A. Ali, M. Elumalai, K. Eluri, S. Srinivasan, J. Acute. Disease., 2013, 316-321.

- [10] N.A. Khalaf, K. Ashok, Shakya, A. Al-Othman, Z. El-agbar, H. Farad, Turk. J. Biol., 2008, 32, 51-55.
- [11] Duffin, W.M. Rollo, I.M. Br, J. Pharm. Col., 1957, 12, 171.
- [12] M.E. Kuehne, E.A. Konopke, J. Med. Chem., 1962, 5, 257.
- [13] J.B. Chylinska, T. Urbanski, J. Med. Chem., 1963, 6, 484.
- [14] L.Y. Hsu, C.H. Lin, Heterocycles., 1996, 43, 2687.
- [15] O.S. Pedersen, E.B. Pedersen, Synthesis., 2000, 2000 (4), 479-495.

Chaitra G et al.

[16] A.J. Cocuzza, D.R. Chidester, B.C. Cordova, S. Jeffrey, R.L. Parsons, Bacheler, Bioorg. Med. Chem. Lett., 2001, 11, 1177.

- [17] T.N. Doan, D.T. Tran., Pharmacol. Pharm., 2011, 2, 282-288.
- [18] C.M. Bhalgat, M.I. Ali, B. Ramesh, G. Ramu, Ara, J. Chem., 2011, 1-8.
- [19] O.V. Singh, H. Han, Tetrahedron Lett., 2007, 48, 2345.
- [20] N. Zonatta, Squizani, A.M.C, Fantinel, N. Bonacorso, J. Braz. Chem. Soc., 2005, 16, 1255-1261.
- [21] Shingare, A.H. Kategaonkar, S.S. Sonar, U. Rajkumar, A.H. Kategaonkar, B. Bapurao, Bull. Korean. Chem. Soc., 2010, 31, 6, 1657.
- [22] Z. Nowakowska, Eur. J. Chem., 2007, 42, 125-137.
- [23] Oyedapo, J. famurewaa, Int. J. P'çog., 1995, 33, 65-69.
- [24] M. Philip, J. Songklanakarin, Scitechnol., 2004, 26, 2, 211-219.
- [25] B.A. Allne, M.M. Michel, A.L. Margareth, Med. Chem., 2014, 4-7.