



## A Review on Antibacterial Activity of Some Isoindole Derivatives

Ferenc Csende\*, Andrea Porkoláb

Taxus Research Laboratory, Bocskai u. 22., H-4080 Hajdúnánás, Hungary

### ABSTRACT

Compounds which are having an isoindole moiety are important classes of the bioactive molecules. Several isoindoles and their fused derivatives were synthesized and evaluated for their efficiency as antimicrobial agents in the past years. However, several novel problems appeared with the treatment of various infections (resistant bacterial or nosocomial infections, multidrug-resistant bacterial, rare and aggressive infections), thus medicinal or pharmaceutical chemists are challenged to discover and find improved novel and more efficient drugs. The aim of this overview is to summarize the most efficient and promising isoindole derivatives to promote these developments of potent and safe drugs with fewer side effects.

**Keywords:** Antibacterial, Enzymes, Inhibition, Isoindoles, Fused rings

### INTRODUCTION

The synthesis, chemistry, structural and pharmacological behaviour of isoindoles were earlier surveyed thoroughly [1]. Isoindoles, constitutional isomers of indoles and their derivatives are less well studied. The isoindole shows structural characteristics indicative of an aromatic system: the carbocyclic ring shows bond alternation reminiscent of that found in naphthalene [2]. Isoindole is much less stable than its isomer indole, thus the saturated, fused and 1- or 1,3-dioxo derivatives are used in the organic and pharmaceutical chemistry practically [3]. The isoindoles are important groups of N-heterocyclic compounds occur widely in living matter and have significant biological roles [4]. Isoindoles constitute the core of many alkaloids, phthalocyanines, synthetic fused heterocycles, isoindoline pigments and isoindole-BODIPY dyes [5-7]. In this overview we could report some isoindole derivatives with regard to their antimicrobial activity. Although most bacteria will not invade other living organism and many more bacteria are harmless to our immune systems or often beneficial, some are pathogenic, with the number of species estimated as fewer than hundred that are seen to cause infectious diseases in humans. People can get infected with pathogenic bacteria by direct (physical) or indirect contacts (water, food, water, wounds and biological products like blood, serum, plasma and airborne transmission). The variety of pathogenic bacteria is quite broad, but there are some that commonly cause infectious diseases. The most common pathogenic bacteria are the *Staphylococcus aureus*, some *Streptococcus* species and certain strains of *Escherichia coli*, they cause also some of the most serious diseases. The cholera is caused by the bacterium *Vibrio cholerae* and usually spreads by drinking infected water and due to unhealthy conditions [8]. There are the main classes and types of the antibacterial drugs: penicillins, aminoglycosides, carbapenems, cephalosporins, glycopeptides, macrolides, polypeptides, quinolones, sulfonamides and finally tetracyclines. The development of new medicines becomes the important and urgent target because of the limited availability of efficient broad-spectrum antibiotic drugs and the increasing evolvement of resistance.

#### 7-Amino- and chloro-isoindole derivatives

Neumann H et al., [9] reported that some *N*-analogue of the corollosporine have notable antibiotic activity, the compounds of which were prepared by the conversion of *N*-methyl-4-aminophthalimides with different equivalents of Grignard reagents. Thus 7-amino-3-hexyl-3-hydroxy-2-methyl-2,3-dihydroisoindol-1-one and its 2,4,6-trimethyl derivative (Figure 1) were found to be active against *Staphylococcus aureus* (inhibition zone: 10 mm).

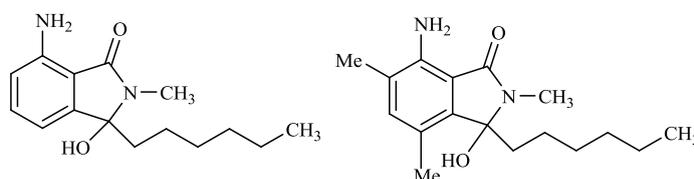


Figure 1: 7-amino-3-hexyl-3-hydroxy-2-methyl-2,3-dihydroisoindol-1-one and its 2,4,6-trimethyl derivative

Some of the chlorinated compounds (Figure 2) showed activity according to the screening results with the minimal inhibitory concentration

(MIC) values of 83.5 and 28.5 µg/ml against *Bacillus subtilis*.

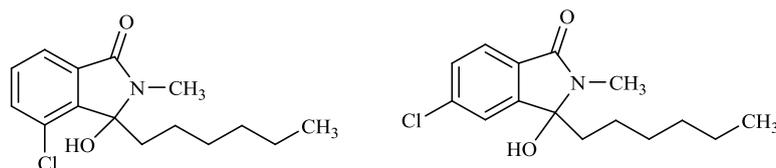


Figure 2: chlorinated compounds

Mikolasch A *et al.*, [10] synthesized further *N*-analogous corollosporines (Figure 3) *via* laccase-catalyzed coupling reaction of the *N*-methyl-4-aminophthalimides with 2,5-dihydroxybenzoic acid derivatives. The obtained products (R = H, Me) were tested in several biological assay to evaluate their antimicrobial effect. According to the results, the compounds prepared inhibited the growth of several gram positive bacterial strains, among them methicillin-resistant *S. aureus* (inhibition zone: 8-10 mm).

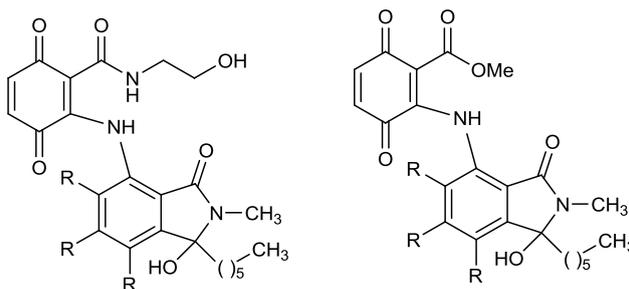


Figure 3: N-analogous corollosporines

#### Tricyclic β-lactams (trinems or tribactams)

The trinems (tribactams) are a novel class of antibiotics, which are tricyclic β-lactams containing a unsaturated or partly saturated isoindole moiety. Heck JV *et al.*, [11] synthesized the fused analogue of the β-lactam Nocardicin A (Figure 4), which is more active against gram negative than gram positive microorganisms. The preparation of the tricyclic cyclonocardicin was achieved starting from 2-bromo-5-hydroxybenzaldehyde in a multi-step procedure. Unfortunately, the isoindole analogue had quite poor stability in neutral aqueous solution and the antibacterial activity was less effective. Christensen BG *et al.*, [12] prepared 6-(1-hydroxyethyl)-cyclonocardicin (Figure 5) by a similar method. This compound is also a novel class of antibiotics, which is bioactive against a broad range of pathogens (*e. g.* *S. aureus*, *E. coli*, *Klebsiella pneumoniae*, *B. subtilis*).

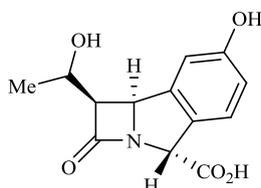


Figure 4: β-lactam Nocardicin

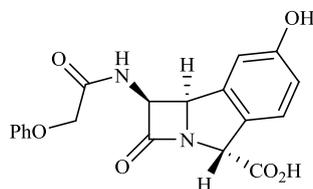


Figure 5: 6-(1-hydroxyethyl)-cyclonocardicin

Hammond SM *et al.*, [13] investigated the *in vitro* activity of hexahydroazeto[2,1-*a*]isoindol-2(1*H*)-one namely, tribactam GV104326 (Figure 6) against gram positive, gram negative and anaerobic bacteria. This compound was extremely active against the Gram-positive bacteria tested, such as *S. aureus* strains and *Staphylococcus epidermidis* were highly sensitive to GV104326.

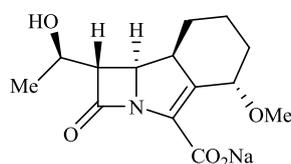


Figure 6: tribactam GV104326

The cumulative MICs were for 90% of the tested strains (MIC90s) ≤ 0.12 µg/ml and MIC90 ≤ 0.06 µg/ml respectively. Among other tested

antibiotics the GV104326 exhibited consistently low MIC90s for *S. aureus*, *E. coli*, *K. pneumoniae*, *Klebsiella oxytoca*, *Proteus mirabilis*, *Proteus vulgaris* and *Clostridium perfringens*. Additional *in vitro* tests and *in vivo* microbiological evaluations of the tricyclic  $\beta$ -lactam GV104326 (Sanfetrinem as sodium salt and Sanfetrinem cilexetil ester, which is a prodrug) were carried out in detail [14-16]. The orally bioavailable and metabolically labile ester derivative (GV118819) is currently undergoing phase II clinical trials. Several methods were developed and patented for the synthesis of GV104326 and its derivatives [17-21]. Andreotti D *et al.*, [22] described the synthesis of all the isomers of 8-methoxy- and 4-methoxy derivatives of compound GV104326 and evaluated their *in vitro* antibacterial activity in comparison with imipenem. According to microbiological test data the (4*S*, 8*S*)-4-methoxy derivative showed the best activities. This research group extended their synthesis and studies to further 4-substituted derivatives of the GV104326 molecule. In these projects Andreotti D *et al.*, [23], on the course of multistep procedure, the 2-(methoxyethoxy)-cyclohex-2-en-1-one (Figure 7, R = Me) or the epoxide derivative of the 4-cyclohexylazetidin-2-one was used as key compound for the construction of the cyclohexane ring (Figure 7, R = OH, F, CN). Compounds (Figure 7) have shown a good activity (MIC 0.03- 8  $\mu$ g/ml) against several bacterial strains.

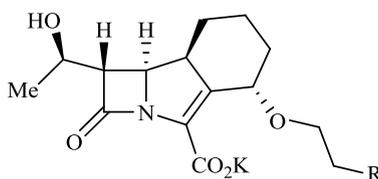


Figure 7: 2-(methoxyethoxy)-cyclohex-2-en-1-one

Fabio RD *et al.*, [24] reported a careful preparation of 2- and 3-aminoethoxy analogue of molecule GV104326. The 2-aminoethoxy derivative was the most active compound (MIC 4 mg/ml) against *Pseudomonas aeruginosa*. Géhanne S *et al.*, [25] achieved synthesis of 4-ureido trinems (Figure 8) from the known amino alcohol intermediate *via* Wittig-type cyclisation and followed by reaction with the corresponding isocyanate. The obtained 4-*N*-methyl-*N'*-alkyl ureido trinems had good antibacterial activity in the course of *in vitro* microbiological tests (MIC  $\leq$  0.12-8  $\mu$ g/ml).

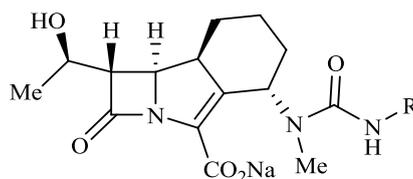


Figure 8: 4-ureido trinems

Kanno O *et al.*, [26] described the synthesis and biological evaluation of the 4-pyrrolidin-3-ylthiomethyl derivative of GV104326 trinem (Figure 9). The authors established that compounds showed potent antimicrobial activity (MIC~1.5  $\mu$ g/ml) against gram positive bacteria including *S. aureus* 535 (MRSA) and higher *in vivo* efficiency against *S. aureus* 507 (MIC 0.78  $\mu$ g/ml).

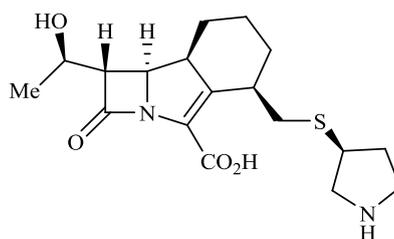


Figure 9: GV104326 trinem

When a bacteria produce  $\beta$ -lactamase enzymes, they hydrolyze the  $\beta$ -lactam ring of the antibiotic, thus may render a drug inactive. The 4-substituted trinems (tribactams) inhibit the broad spectrum  $\beta$ -lactamase enzymes and these compounds have different antibiotic activity generally. Copar A *et al.*, [27] reported design, synthesis and pharmaceutical evaluation of novel unsubstituted trinems and their 6-thia derivatives (Figure 10, X = CH<sub>2</sub> or S). Some products showed activity at the enzyme  $\beta$ -lactamase I from *Bacillus cereus* and another class A  $\beta$ -lactamase (*E. coli* TEM1). 38-61% Inhibition of  $\beta$ -lactam hydrolysis was observed in 100  $\mu$ mol/l of lactamase enzyme I from *B. cereus*.

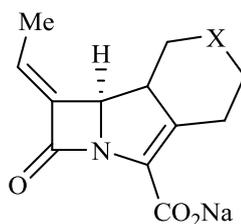


Figure 10: Unsubstituted trinems and their 6-thia derivatives

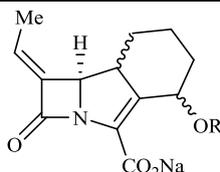


Figure 11: stereoisomeric 4(S or R)-alkoxy trimems

Plantan I *et al.*, [28] described the structure-based design, synthesis and biological activity of several stereoisomeric 4(*S* or *R*)-alkoxy trimems (Figure 11), which are analogue of the lead compound ( $R = 4(S)\text{-OMe}$ , namely LK-157). The  $\beta$ -lactamase inhibitory activity of the derivatives (OEt, OPr, OBU, *etc.*) compared to LK-157 are decreased despite various lipophilicity. When Paukner S *et al.*, [29] tested LK-157 against the purified class A (TEM-1 and SHV-1)  $\beta$ -lactamases, ~50% inhibitory concentrations ( $IC_{50} \sim 55$  and  $\sim 151$  nM) were observed, furthermore the inhibition of the AmpC enzyme was significant on average ( $IC_{50} \sim 62$  nM).

#### Isoindolinyl-4-oxoquinoline-3-carboxylic acids

Besides tricyclic  $\beta$ -lactams the quinolone-carboxylic acid derivatives represent important members of the large group of broad-spectrum active antibacterial drugs. These are active against both gram positive and gram negative bacteria and the compounds function by inhibiting DNA gyrase and the topoisomerase IV, which is one of two the type-II topoisomerases in bacteria. Todo Y *et al.*, [30,31] patented an invention related to the synthesis of bioactive 7-isoindolinyl-quinolone-carboxylic acid derivatives, where a 6-nonfluorinated 1-cyclopropyl-8-(difluoromethoxy)-7-(1-methylisoindolin-5-yl)-4-oxoquinoline-3-carboxylic acid (formally T-3811 and BMS-284756 or Garenoxacin) (Figure 12) was found as the candidate compound.

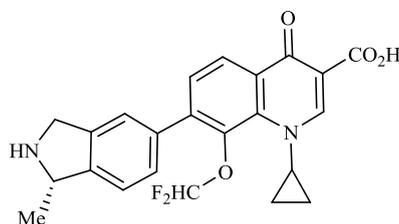


Figure 12: Structure of Garenoxacin

General synthetic route to 1-cyclopropyl-7-(isoindolin-5-yl)-4-oxoquinoline-3-carboxylic acids was achieved *via* Stille-type coupling reaction between *N*-Cbz-1-methyl-5-(tributylstannyl)isoindoline and ethyl 7-bromo-1-cyclopropyl-4-oxoquinoline-3-carboxylate was described by Reuman M *et al.*, [32]. The alternative method was also reported by Todo Y *et al.*, in patents [30,31] where the inventors applied the Suzuki cross-coupling reaction of the 7-[1-methyl-2-(triphenylmethyl)isoindolin-5-yl]boronic acid with ethyl 7-bromo-1-cyclopropyl-4-oxoquinoline-3-carboxylate. Takahata M *et al.*, [33] investigated this so-called des-F(6)-quinolone compound by *in vitro* and *in vivo* methods for evaluation of antibacterial activity in comparison to other fluoroquinolones (ciprofloxacin, levofloxacin, trovafloxacin). T-3811 exhibited potent activity against *S. aureus* (MIC 0.025  $\mu\text{g/ml}$ ), the methicillin-resistant *Staphylococcus epidermidis* (MIC 6.25  $\mu\text{g/ml}$ ), *S. pneumoniae* (MIC 0.05  $\mu\text{g/ml}$ ) and against *M. tuberculosis* was exceptionally effective (MIC 0.0625  $\mu\text{g/ml}$ ). In the course of *in vitro* studies of BMS-284756 (T-3811) Rhomberg PR *et al.*, [34] observed high activity against *Campylobacter jejuni*, *Helicobacter pylori*, *Legionella spp.* (MIC<sub>50</sub> 0.032; 0.004 and 0.25  $\mu\text{g/ml}$ ) micro aerophilic and anaerobic bacteria. The results demonstrated that BMS-284756 had comparable or greater activity than other investigated quinolones. Hayashi K *et al.*, [35] described the synthesis of a series of 1-cyclopropyl-7-(isoindolin-5-yl)-4-oxoquinoline-3-carboxylic acids (Figure 13), their antibacterial and toxicological evaluation. The various substituents of the compounds prepared are  $R_1, R_2 = \text{H}$  or/and  $\text{CH}_3$ ,  $R_3 = \text{H}, \text{OCH}_3, \text{OCHF}_2$ , while  $R_4 = \text{H}$  or  $\text{F}$  and the synthetic procedures were similar to the methods, that Todo Y *et al.*, described [30,31].

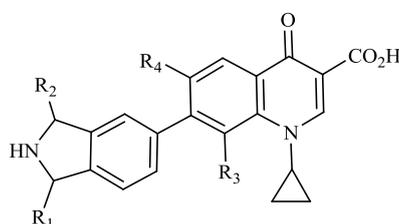


Figure 13: Structure of 1-cyclopropyl-7-(isoindolin-5-yl)-4-oxoquinoline-3-carboxylic acids

Earlier Yatsunami T *et al.*, [36] reported the synthesis and evaluation of antibiotic effects of several 7-(2-isoindolinyl)-1-cyclopropyl-6,8-difluoro-1,4-dihydro-4-oxoquinoline-3-carboxylic acid derivatives (Figure 14). Generally, the reaction of 6,7,8-trifluoro-dihydro-4-oxoquinoline-3-carboxylic acids with the corresponding isoindoline in DMF at 120°C for 1.5 h gave the desired products in good yields. This compound showed remarkable antibacterial activity against *S. aureus*, *E. coli* and *P. aeruginosa* (MIC <0.025  $\mu\text{g/ml}$ ; 0.2  $\mu\text{g/ml}$  and 0.39  $\mu\text{g/ml}$ ). Similarly, Petersen U *et al.*, [37] synthesized the tetra-hydro-isoindolinyl analogue (Figure 15), which according to the invention also exhibits surprising increases in action against the above-mentioned bacteria and *Enterococcus faecalis*.

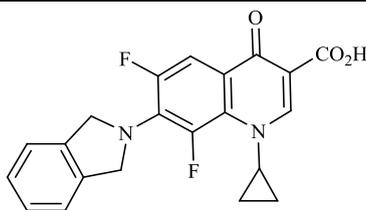


Figure 14: Structure of 7-(2-isoindolinyl)-1-cyclopropyl-6,8-difluoro-1,4-dihydro-4-oxoquinoline-3-carboxylic acid derivatives

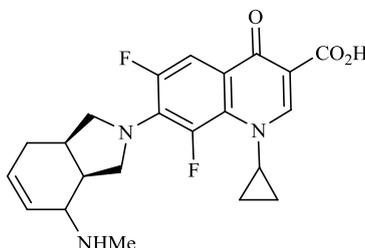


Figure 15: Structure of tetrahydroisoindolyl analogue

### Isoindolinone and its fused heterocycles

Mohamed EK *et al.*, [38] synthesized some novel condensed pyrimidine derivatives using the 3-amino-1,9-dioxo-1,9-dihydropyrimido[6,1-*a*]isoindole-4-carbonitrile as a key intermediate. Thus tetracyclic pyrimido[6,1-*a*]isoindole-2-carboxylate (Figure 16) was prepared by the reaction of this amino-carbonitrile with diethyl malonate. The condensation of the key compound nitrile with triethyl orthoformate in refluxing acetic anhydride gave pyrimido[6,1-*a*]isoindolylimidoformate (Figure 17) and the reaction of aminocarbonitrile with 1,2-ethylenediamine in the presence of carbon disulfide afforded 4-(imidazol-2-yl)pyrimido[6,1-*a*]isoindole-1,9-dione (Figure 18).

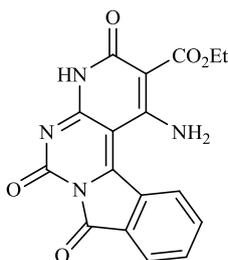


Figure 16: Structure of tetracyclic pyrimido[6,1-*a*]isoindole-2-carboxylate

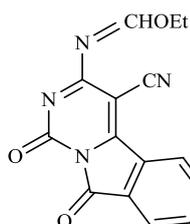
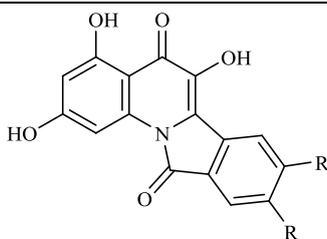


Figure 17: Structure of pyrimido[6,1-*a*]isoindolylimidoformate



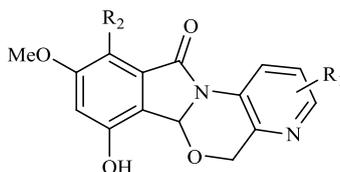
Figure 18: Structure of 4-(imidazol-2-yl)pyrimido[6,1-*a*]isoindole-1,9-dione

The preliminary bioactivity screening of these compounds was performed at 50 µg/ml against *S. aureus* and *E. coli* and results showed that imidazolyl derivative (Figure 18) and imidoformate derivative (Figure 17) had significant antimicrobial activities, while the tetracyclic isoindolone derivative (Figure 16) and the imidoformate derivative (Figure 17) had medium activity. Sui Z *et al.*, [39] described synthesis of a series of isoindolo[2,1-*a*]quinolines from the reaction of substituted phthalic anhydrides with 2-aminoacetophenones in refluxing xylene. Demethylation of tetracyclic compounds was carried out with pyridine hydrochloride resulting in potent DNA-gyrase and topoisomerase-II inhibitor agents (Figure 19).

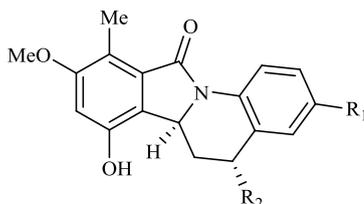


**Figure 19: Demethylation of tetracyclic compounds**

When substituents were changed to dichloro or dihydroxy ( $R = Cl, OH$ ) on the isoindolone moiety, the compounds showed very good activity in bacterial DNA-gyrase assay ( $IC_{50} = 82 \mu g/ml$  and  $22 \mu g/ml$ ) and in human topoisomerase II assay ( $IC_{50} = 13 \mu g/ml$  and  $0.29 \mu g/ml$ ), thus some of them are more potent than ellipticine or nalidixic acid. Lübbers T *et al.*, [40] reported design and simple synthesis of isoindolo[2,1-*a*]quinolinones (Figure 20;  $R_1 = Cl, R_2 = OH$  or  $R_1 = OMe, R_2 = H$ ) and isoindolinone fused 1,4-dihydro-2*H*-pyrido[3,2-*d*][1,3]oxazines (Figure 21;  $R_1 = H, R_2 = Me$  or  $R_1 = 5,6-(CH_2)_4, R_2 = H$ ). Latter compounds (Figure 21) were proved to be effective DNA gyrase inhibitor in the supercoiling assay *in vitro E. coli* tests, the maximal non-effective concentration was low (MNEC =  $0.13 \mu g/ml$ ).

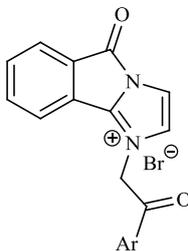


**Figure 20: Structure of pyrido[3,2-*d*][1,3]oxazine derivatives**



**Figure 21: Structure of isoindolo[2,1-*a*]quinolinones**

The *in vitro* antibiotic activities of pyrido[3,2-*d*][1,3]oxazine derivatives (Figure 20) were remarkable against *S. aureus* strains (MIC =  $0.5-2.0 \mu g/ml$ ), while isoindolo[2,1-*a*]quinolinones (Figure 21) had moderate activity against same bacteria. Cyclo-condensation reaction of methyl 2-formylbenzoate with various anilines under basic conditions and subsequent addition of the obtained amination to thiols or electron rich heteroarenes, including furans, indoles resulted in novel 2,3-dihydroisoindolones. The allyl, 2-oxopropanyl, formylmethyl and hydroxyethyl derivatives were also prepared in further reactions, however these compounds showed moderate DNA gyrase inhibition or were almost inactive. These isoindolones led to the thorough SAR study of the novel phenolic DNA gyrase inhibitors. Narsimha S *et al.*, [41] prepared a series of the imidazo[2,1-*a*] isoindolium bromide derivatives (Figure 22) from 2-iodobenzoic acid and *N,N*-carbonyldiimidazole (CDI) in a one-pot palladium-catalyzed coupling reaction in good yields.



**Figure 22: Structure of imidazo[2,1-*a*]isoindolium bromide derivatives**

All the synthesized compounds were screened for their *in vitro* antibacterial activity and two derivatives showed excellent inhibition against both gram positive and gram negative bacteria (Table 1).

**Table 1: *In vitro* antibacterial activity (MIC in  $\mu g/ml$ ) of 5*H*-imidazo[2,1-*a*]isoindolium bromides**

Compound	<i>Staphylococcus aureus</i>	<i>Bacillus subtilis</i>	<i>Escherchia coli</i>	<i>Proteus vulgaris</i>
1 (Ar = 4-Me-Ph-)	3.125	25	12.5	6.25
2 (Ar = 4-Ph-Ph-)	3.125	3.125	12.5	6.25
3 Streptomycin	6.25	6.25	6.25	3.125

MIC: Minimum Inhibitory Concentration

Earlier Breitenbach JC *et al.*, [42] reported the synthesis and *in vitro* antibacterial activity of a range of N-substituted isoindolin-5-ones (Figure 23; R= Me, Pri, Bui, Ph, etc). The tested compounds were prepared by refluxing  $\alpha$ -amino acids (e. g. stereoisomers of alanine, valine, serine and aspartic acid) with *o*-phthalaldehyde under mild and simple reaction conditions.

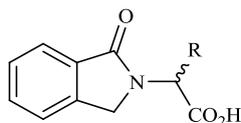


Figure 23: Structure of Isoindolinones

Isoindolinones (Figure 23) exhibited promising antibacterial activity (MIC 0.328-3.6 mg/ml) against *B. subtilis*, *S. aureus*, *Micrococcus roseus* and *E. coli* bacteria similar to  $\beta$ -lactam antibiotics. The carboxyl functional group in compounds seems to be necessary for antibacterial activity.

### CONCLUSION

Over the past few decades numerous isoindolines and their fused derivatives were developed, synthesized and tested for antibacterial activities and the inhibition of pathogen bacteria reproduction. The mechanism of action of the synthesized and tested compounds was usually different, but sometimes the mechanism is complex or unknown so far. It may refer to inhibition of cell growth by interaction and modulation of a direct bio molecular target, such as a specific protein (enzyme, receptor) or nucleic acid (DNA). Hopefully, these results will serve support the medicinal chemists, researchers to design, coordinate and accomplish new approaches towards discovery of novel drugs.

### ACKNOWLEDGEMENT

The authors would like to thank G. Csende for his careful revision of manuscript and his useful remarks and we also would like to express our thanks to the Institute of Pharmaceutical Chemistry, University of Szeged for the professional help.

### REFERENCES

- [1] F. Csende, F. Miklós, G. Stájer, *Curr. Org. Chem.*, **2012**, 16(8), 1005, and references therein.
- [2] R. Bonnet, M.B. Hursthouse, S.A. North, J. Trotter, *J. Chem. Soc., Perkin Trans. 2*, **1985**, 293.
- [3] R.K. Bhatia, *Curr. Top. Med. Chem.*, **2017**, 17(2), 189.
- [4] T.S.A. Heugebaert, B.I. Roman, C.V. Stevens, *Chem. Soc. Rev.*, **2012**, 41, 5626.
- [5] C. Yu, Q. Wu, J. Wang, Y. Wei, E. Hao, L. Jiao, *J. Org. Chem.*, **2016**, 81(9), 3761.
- [6] Y. Ni, J. Wu, *Org. Biomol. Chem.*, **2014**, 12, 3774.
- [7] H. Liu, Y. Wu, Z. Li, H. Lu, *J. Porphyrins Phthalocyanines*, **2014**, 18, 679.
- [8] C.J.L. Murray, *Lancet*, **2016**, 388, 1459.
- [9] H. Neumann, D. Strübink, M. Lalk, S. Klaus, S. Hübner, A. Spannenberg, U. Lindequist, M. Beller, *Org. Biomol. Chem.*, **2006**, 4, 1365.
- [10] A. Mikolasch, S. Hessel, M.G. Salazar, H. Neumann, K. Manda, D. Gördes, E. Schmidt, K. Thurow, E. Hammer, U. Lindequist, M. Beller, F. Schauer, *Chem. Pharm. Bull.*, **2008**, 56(6), 781.
- [11] J.V. Heck, B. G. Christensen, *Tetrahedron Lett.*, **1981**, 22(50), 5027.
- [12] B.G. Christensen, J.V. Heck, M. J. Szymonifka, US Patent 4, 374, 848, **1983**.
- [13] E. Di Modugno, I. Erbeti, L. Ferrari, G. Galassi, S.M. Hammond, L. Xerri, *Antimicrob. Agents Chemother.*, **1994**, 38(10), 2362.
- [14] R. Wise, J.M. Andrews, N. Brenwald, *Antimicrob. Agents Chemother.*, **1996**, 40(5), 1248.
- [15] K.V. Singh, T.M. Coque, B.E. Murray, *Antimicrob. Agents Chemother.*, **1996**, 40(9), 2142.
- [16] S. Tamura, S. Miyazaki, K. Tateda, A. Ohno, Y. Ishii, T. Matsumoto, N. Furuya, K. Yamaguchi, *Antimicrob. Agents Chemother.*, **1998**, 42(7), 1858.
- [17] B. Tamburini, A. Perboni, T. Rossi, D. Donati, D. Andreotti, G. Gaviraghi, R. Carlesso, C. Bismara, EP Patent 0416 953 (**1991**).
- [18] M. Sendai, T. Miwa, EP Patent 0422 596 (**1995**).
- [19] S. Michiyuki, T. Suita, US Patent 5, 393, 751 (**1995**).
- [20] M. Sendai, T. Miwa, US Patent 5, 459, 260 (**1995**).
- [21] I. Plantan, A. Prezelj, U. Urleb, B. Mohar, M. Stephan, EP Patent 2135 871 (**2009**).
- [22] D. Andreotti, T. Rossi, G. Gaviraghi, D. Donati, C. Marchioro, E. Di Modugno, A. Perboni, *Bioorg. Med. Chem. Lett.*, **1996**, 6(4), 491.
- [23] D. Andreotti, S. Biondi, R. Di Fabio, D. Donati, E. Piga, T. Rossi, *Bioorg. Med. Chem. Lett.*, **1996**, 6(16), 2019.
- [24] R. Di Fabio, D. Andreotti, S. Biondi, G. Gaviraghi, T. Rossi, *Bioorg. Med. Chem. Lett.*, **1996**, 6(16), 2025.
- [25] S. Géhanne, E. Piga, D. Andreotti, S. Biondi, D. Pizzi, *Bioorg. Med. Chem. Lett.*, **1996**, 6(22), 2791.
- [26] O. Kanno, Y. Shimoji, S. Ohya, I. Kawamoto, *J. Antibiot.*, **2000**, 53(4), 404.
- [27] A. Copar, T. Prevec, B. Anžič, T. Mesar, L. Selič, M. Vilar, T. Solmajer, *Bioorg. Med. Chem. Lett.*, **2002**, 12(6), 971.
- [28] I. Plantan, L. Selič, T. Mesar, P. Š. Anderluh, M. Oblak, A. Prezelj, L. Hesse, M. Andrejašič, M. Vilar, D. Turk, A. Kocijan, T. Prevec, G. Vilfan, D. Kocjan, A. Čopar, U. Urleb, T. Solmajer, *J. Med. Chem.*, **2007**, 50(17), 4113.
- [29] S. Paukner, L. Hesse, A. Prezelj, T. Šolmajer, U. Urleb, *Antimicrob. Agents Chemother.*, **2009**, 53(2), 505.
- [30] Y. Todo, K. Hayashi, M. Takahata, Y. Watanabe, H. Narita, WO Patent 029 102 (**1997**).
- [31] Y. Todo, K. Hayashi, M. Takahata, Y. Watanabe, H. Narita, US Patent 6, 025, 370 (**2000**).
- [32] M. Reuman, S.J. Daum, B. Singh, M.P. Wentland, R.B. Perni, P. Pennock, P.M. Carabateas, M.D. Gruett, M.T. Saindane, P.H. Dorff, S.A. Coughlin, D.M. Sedlock, J.B. Rake, G.Y. Leshner, *J. Med. Chem.*, **1995**, 38(14), 2531.
- [33] M. Takahata, Y. Yamashiro, M. Yonezawa, H. Araki, Y. Todo, S. Minami, Y. Watanabe, H. Narita, *Antimicrob. Agents Chemother.*, **1999**, 43(5), 1077.
- [34] P.R. Rhombert, D.J. Biedenbach, R.N. Jones, *Diagn. Microbiol. Infect. Dis.*, **2001**, 40(1-2), 45.
- [35] K. Hayashi, M. Takahata, Y. Kawamura, Y. Todo, *Arzneim.-Forsch./Drug Res.*, **2002**, 52(12), 903.
- [36] T. Yatsunami, A. Yazaki, S. Inoue, H. Yamamoto, M. Yokomoto, Y. Nomiya, S. Noda, EP Patent 0343 560 (**1989**).

- [37] U. Petersen, A. Krebs, T. Schenke, F. Kunisch, T. Philipps, K. Grohe, K.D. Bremm, R. Endermann, K.-G. Metzger, I. Haller, H.J. Zeiler, US Patent, **1995**, 464, 796
- [38] E.K. Mohamed, W.S. Shehab, *J. Korean Chem. Soc.*, **2011**, 55(6), 988.
- [39] Z. Sui, J. Altom, V.N. Nguyen, J. Fernandez, J.I. Bernstein, J.J. Hiliard, J.F. Baret, B.L. Podlogar, K. A. Ohemeng, *Bioorg. Med. Chem.*, **1998**, 6(6), 735.
- [40] T. Lübbers, P. Angehrn, H. Gmünder, S. Herzig, *Bioorg. Med. Chem. Lett.*, **2007**, 17(16), 4708.
- [41] S. Narsimha, K. Battula, N.V. Reddy, *Synth. Commun.*, **2017**, 47(9), 928.
- [42] J.C. Breytenbach, S. van Dyk, I. van den Heever, S.M. Allin, C. C. Hodkinson, C.J. Northfield, M.I. Page, *Bioorg. Med. Chem. Lett.*, **2000**, 10(15), 1629.