



Scholars Research Library

Der Pharma Chemica, 2012, 4(4):1626-1630  
(<http://derpharmachemica.com/archive.html>)



ISSN 0975-413X  
CODEN (USA): PCHHAX

## Antiglycation and anticancer activity of some newer synthetic flavones

Jayashree B. S<sup>1\*</sup>, Piyush Chaturvedi<sup>1</sup>, Venkatachalam H<sup>2</sup>, P V R Chaudary<sup>1</sup>, Yogendra Nayak<sup>3</sup>

<sup>1</sup>Department of Pharmaceutical Chemistry, Manipal College of Pharmaceutical Sciences, Manipal

<sup>2</sup>Department of Chemistry, KMC International Center, Manipal University, Manipal

<sup>3</sup>Department of Pharmacology, Manipal College of Pharmaceutical Sciences, Manipal

---

### ABSTRACT

Flavonoids have been used for prevention and treatment for diabetes and cancer. Several attempts have been made earlier to substitute the  $\gamma$ -benzopyrone pharmacophore in the flavonoid ring. We have substituted  $\alpha$ -position in the  $\gamma$ -benzopyrone ring with a thiophenyl group. Thus the newer chalcones (JPC-1 and JPC-19), 3-hydroxy flavones (JPC-2 and JPC-20) and many new substituted 3-benzyloxy derivatives of flavones were synthesized, purified, characterized by their spectral studies, and their  $\log P$ ,  $\epsilon_{\max}$  values were determined. The synthesized compounds were screened for *in vitro* antiglycation and anticancer activities. In the *in vitro* glycosylation using haemoglobin-glucose method, test compounds JPC-6 and JPC-10 showed activity less than 100  $\mu\text{g/ml}$  when compared to that of  $\alpha$ -tocopherol acetate. All the other test compounds showed antiglycation activity more than 100  $\mu\text{g/ml}$ . In the BSA-glucose assay, JPC-8 showed antiglycation activity less than 100  $\mu\text{g/ml}$  and was comparable to quercetin. Test compounds JPC-10 and JPC-13 showed significant *in vitro* anticancer activity with  $\text{IC}_{50}$  values 43.50 and 67.44  $\mu\text{g/ml}$  respectively, when they were tested on Ehrlich's Ascitic Carcinoma (EAC) cells by trypan blue exclusion assay.

**Key words:** Antiglycation, Anticancer, Flavones, Chalcones

---

### INTRODUCTION

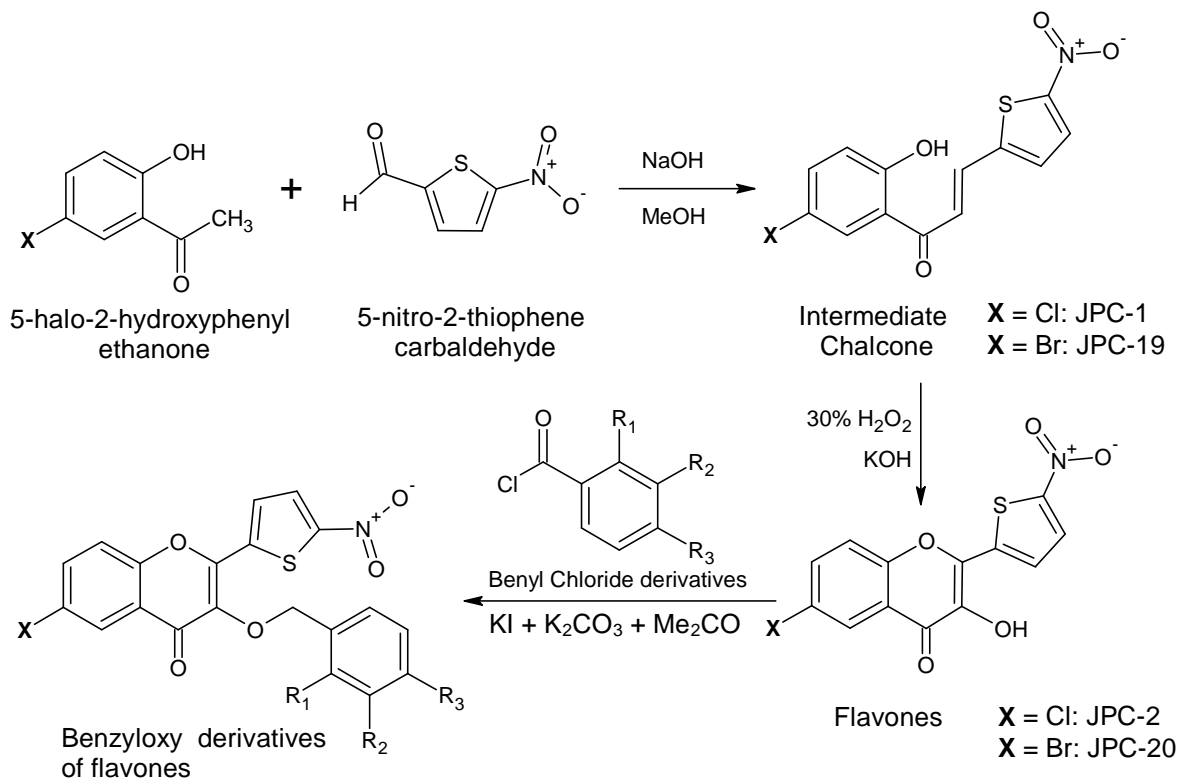
The oxidative stress plays an important role in pathogenesis and progression of diabetes mellitus, cancer and related metabolic complications[1]. Free radicals are formed disproportionately in diabetes by increased glucose oxidation, nonenzymatic glycation of proteins, and the subsequent oxidative degradation of glycated proteins can lead to damage of cellular organelles and enzymes, increased lipid peroxidation, and development of insulin resistance[2]. Flavonoids have been reported for antidiabetic and anticancer activity[3-4]. Some of the flavonoids were reported for their antiglycation activity whereby they have ameliorated diabetes and prevented diabetes related metabolic complications[5]. Similarly, flavonoids have been exploited for the treatment and prevention of cancer and one of the flavonoids, flavopiridol was approved for therapy in leukemia where, it acts by inhibiting CDKs in leukemia cells[6].

All flavonoids have  $\gamma$ -benzopyrone as a pharmacophore with phenyl ring attached at  $\alpha$  or  $\beta$ -position[7]. Earlier we have reported several such synthetic flavonoids and their biological activity[8-10]. In the present study, we have introduced thiophenyl moiety at  $\alpha$ -position of  $\gamma$ -benzopyrone, to study the enhanced biological activity. The newly synthesized flavonoids are further screened for their anti-glycation and anticancer activity.

## MATERIALS AND METHODS

## Chemicals and instruments

Chemicals were purchased from Aldrich, Himedia, Merc and Rankem. All the chemicals were of AR and LR grade and solvents were of HPLC grade. Melting points were determined on a melting point apparatus (Shital Scientific Industries, Mumbai) and are uncorrected. The reactions were monitored by TLC and the R<sub>f</sub> values were determined on pre-coated TLC plates. λ<sub>max</sub> and ε<sub>max</sub> for the test compounds were obtained on UV-visible spectrophotometer (Shimadzu UV-Visible spectrophotometer UV-1650 PC) in methanol (HPLC grade). The FTIR studies were done on Shimadzu FTIR 8310. <sup>1</sup>H NMR spectra were taken on a NMR (AMX 400) and the mass spectra were recorded on QP5050-GC-MS (Shimadzu, Japan). LogP values for the test compounds were determined using shake flask method [9].



Compound	X	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>	Compound	X	R <sub>1</sub>	R <sub>2</sub>	R <sub>3</sub>
JPC-3	-Cl	-H	-H	-NO <sub>2</sub>	JPC-11	-Cl	-H	-H	-F
JPC-4	-Cl	-NO <sub>2</sub>	-H	-H	JPC-12	-Cl	-F	-H	-H
JPC-5	-Cl	-H	-H	-Cl	JPC-13	-Br	-H	-H	-NO <sub>2</sub>
JPC-6	-Cl	-H	-NO <sub>2</sub>	-H	JPC-14	-Br	-H	-NO <sub>2</sub>	-H
JPC-7	-Cl	-Cl	-H	-H	JPC-15	-Br	-NO <sub>2</sub>	-H	-H
JPC-8	-Cl	-H	-H	-OCH <sub>3</sub>	JPC-16	-Br	-H	-H	-Cl
JPC-9	-Cl	-H	-OCH <sub>3</sub>	-H	JPC-17	-Br	-Cl	-H	-H
JPC-10	-Cl	-H	-H	-CH <sub>3</sub>	JPC-18	-Br	-H	-H	-F

Fig.1. Scheme of synthesis for chalcones and flavones

## General procedure for the synthesis

Equi-molar concentrations of 5-halo-2-hydroxyphenyl ethanone and 5-nitro-2-thiophene carbaldehyde were stirred in an ice-cold condition in the presence of NaOH for 5 hours and it was then poured into an ice cold HCl, which gave intermediate chalcones (JPC-1 and JPC-19; Fig.1). The Intermediate chalcones were then purified by recrystallization and were further used for the preparation of the corresponding 3-hydroxyflavones. The intermediate

chalcones was dissolved in methanol and NaOH. Further, resulting solution was cooled and stirred at ice-cold condition with drop-wise addition of 30% H<sub>2</sub>O<sub>2</sub>. The final solution was stirred for 5 hours and the mixture was then poured onto ice-cold HCl to get the corresponding 3-hydroxyflavone derivative (JPC-2 and JPC-20; Fig. 1).

To a suspension of the 3-hydroxyflavone derivative, benzyl halides, KI, and freshly ignited anhydrous K<sub>2</sub>CO<sub>3</sub>, dry acetone was added and refluxed for 5 hours. The reaction mixture was then filtered, evaporated and was subjected to percolation by passing through column of silica-gel to obtain corresponding flavone derivatives (JPC-3 to JPC-18). All the flavones and chalcones were then purified by recrystallization and they were obtained in high purity and the structure was later confirmed by melting point, UV, IR, Mass and NMR spectral studies.

## BIOLOGICAL ACTIVITY

### Antiglycation activity

The bovine serum albumin (BSA)-glucose assay was carried out in order to quantify anti-glycosylation activity of test compounds [8]. The inhibition of glycosylation by test compounds was calculated and expressed as IC<sub>50</sub> with quercetin as standards.

In hemoglobin-glucose assay, solution of test compounds at different concentrations was incubated in a mixture containing glucose (2%), hemoglobin (0.6%) and gentamycin (0.2%) in 10 mM phosphate buffer (pH 7.4). The reaction mixtures were then incubated in dark at room temperature for 72 hours with intermittent shaking and glycosylated hemoglobin was determined using NBT-reagent [11]. The antihemoglobin-glucose glycation was determined using  $\alpha$ -tocopherol acetate as standard.

### Anticancer activity by trypan blue exclusion assay

The cancer cell lines [Ehrlich's Ascitic Carcinoma (EAC)], were obtained from National Centre for Cell Sciences (NCCS), Pune, and cultured at the cell lab facilities of Department of Pharmacology, Manipal College of Pharmaceutical Sciences and further, they were maintained as ascitis tumor by serial transplantation in Swiss albino mice. The ascitic fluid was withdrawn from the peritoneal fluid, washed with PBS and cell viability was then checked by trypan blue dye using haemocytometer. The stock cell suspension of 1 X 10<sup>4</sup> was made with phosphate buffer saline (PBS) from which, 0.1 ml of the suspension was taken in sterile test tubes and incubated with 0.1 ml of varying concentrations of test compounds in 0.1% DMSO and 0.7 ml of PBS for 3 hours at 37 °C [12]. After incubation, 0.1 ml of 1% w/v trypan blue solution was added and mixed well. The total number of dead and living cells was then counted using haemocytometer and the percentage viability was calculated as follows:

Percentage viable cells = (Number of unstained cells / Total number of cells) × 100

Table 1: Showing physicochemical properties of different substituted flavone \*

Comp Code	MF	% Yield	MP(°C)	Rf <sup>#</sup>	$\lambda_{\max}$	$\epsilon_{\max}$ (l mol/cm)	logP
JPC-1	C <sub>13</sub> H <sub>8</sub> ClNO <sub>4</sub> S	56	68	0.46	372	23687.48	2.84
JPC-2	C <sub>13</sub> H <sub>6</sub> ClNO <sub>5</sub> S	47	86	0.35	295	14852.44	2.38
JPC-3	C <sub>20</sub> H <sub>11</sub> ClN <sub>2</sub> O <sub>7</sub> S	39	106	0.86	271	1191.61	1.78
JPC-4	C <sub>20</sub> H <sub>11</sub> ClN <sub>2</sub> O <sub>7</sub> S	34	116	0.87	264	11162.08	1.96
JPC-5	C <sub>20</sub> H <sub>11</sub> Cl <sub>2</sub> NO <sub>5</sub> S	25	104	0.51	260	10190.28	1.29
JPC-6	C <sub>20</sub> H <sub>11</sub> ClNO <sub>5</sub> S	28	112	0.89	289	13024.00	1.58
JPC-7	C <sub>20</sub> H <sub>11</sub> Cl <sub>2</sub> NO <sub>5</sub> S	29	102	0.55	259	10816.42	1.19
JPC-8	C <sub>21</sub> H <sub>14</sub> ClNO <sub>6</sub> S	28	106	0.43	280	12461.58	1.64
JPC-9	C <sub>21</sub> H <sub>14</sub> ClNO <sub>6</sub> S	30	110	0.47	275	11046.31	1.76
JPC-10	C <sub>21</sub> H <sub>14</sub> ClNO <sub>5</sub> S	35	108	0.59	262	10238.30	2.75
JPC-11	C <sub>20</sub> H <sub>11</sub> ClFNO <sub>5</sub> S	31	104	0.39	263	10270.06	1.9
JPC-12	C <sub>20</sub> H <sub>11</sub> ClFNO <sub>5</sub> S	33	104	0.37	264	10540.09	1.95
JPC-13	C <sub>20</sub> H <sub>11</sub> BrN <sub>2</sub> O <sub>7</sub> S	37	118	0.72	284	13721.64	1.78
JPC-14	C <sub>20</sub> H <sub>11</sub> BrN <sub>2</sub> O <sub>7</sub> S	35	122	0.77	270	12656.08	-
JPC-15	C <sub>20</sub> H <sub>11</sub> BrN <sub>2</sub> O <sub>7</sub> S	33	114	0.73	290	14127.29	-
JPC-16	C <sub>20</sub> H <sub>11</sub> BrClNO <sub>5</sub> S	32	132	0.42	264	10538.01	-
JPC-17	C <sub>20</sub> H <sub>11</sub> BrClNO <sub>5</sub> S	31	128	0.44	260	11805.46	-
JPC-18	C <sub>20</sub> H <sub>11</sub> BrFNO <sub>5</sub> S	28	130	0.42	265	11728.24	-
JPC-19	C <sub>13</sub> H <sub>8</sub> BrNO <sub>4</sub> S	54	76	0.43	369	22578.31	-
JPC-20	C <sub>20</sub> H <sub>8</sub> BrNO <sub>4</sub> S	44	90	0.54	285	15346.92	-

\* Solvents used for recrystallization of the test compounds was methanol except for the compounds JPC-1, JPC-2, JPC-19 and JPC-20, it was glacial acetic acid.

<sup>#</sup> Solvent system: n-hexane: ethyl acetate (6:2) for JPC-1, JPC-2, JPC-19, JPC-20; chloroform: ethyl acetate (7:3) for JPC-3, JPC-4, JPC-5, JPC-6, JPC-8, JPC-9, JPC-10, JPC-13, JPC-14; chloroform: acetone (7:3) for JPC-7, JPC-11, JPC-12, JPC-16, JPC-17, JPC-18.

## RESULTS AND DISCUSSION

**Chemistry**

The newer chalcones and 3-hydroxy flavones were synthesized with yield, ranging from 25 to 55%. The logP values were in the range between 1.5 and 3 as shown in the table 1.

**Spectral data for the representative molecule**

**6-chloro-3-[(4-methoxybenyl)oxy]-2-(5-nitrothiophen-2-yl)-4H-chromen-4-one (JPC-8):** IR (KBr) (cm<sup>-1</sup>): 1645.33 (C-O-CH<sub>2</sub>), 1502.26 [Ar-NO<sub>2</sub> (N=O)], 752 (C-Cl); <sup>1</sup>HNMR (DMSO-d<sub>6</sub>): δ 3.72 (s 12), δ 0.0082 (s 3); MS: m/z 444 (M<sup>+</sup>).

**BIOLOGICAL ACTIVITY**

**Antiglycation activity:** The test compounds were subjected for their antiglycation activity and compared with the standard quercetin. In BSA-glucose antiglycation assay, the synthetic compounds JPC-3, JPC-8, JPC-10 and JPC-12 showed IC<sub>50</sub> values at 340, 30, 170 and 229 µg/ml respectively as shown in table 2 and were significantly higher than that of the standard quercetin (IC<sub>50</sub> = 354 µg/ml). Further, in haemoglobin-glucose assay JPC-6 and JPC-10 showed considerable activity and the IC<sub>50</sub> was less than 100 µg/ml, compared to that of the standard α-tocopherol acetate (IC<sub>50</sub>: 12.7 µg/ml). All remaining test compounds showed antiglycation activity more than 100 µg/ml as shown in table 2.

**Anticancer activity:** The test compounds synthesized were subjected for their anticancer activity by trypan blue dye exclusion assay using EAC cells. Trypan blue dye exclusion assay was performed to assess the preliminary anticancer activity of the synthesized compounds. Test Compounds, JPC-10 and JPC-13 were found to be cytotoxic with IC<sub>50</sub> values at 43.50 and 67.44 µg/ml. However, test compounds, JPC-5, JPC-7, JPC-11 and JPC-12 showed anticancer activity with IC<sub>50</sub> values of 151.50, 132.49, 128.22 and 147.45 µg/ml respectively, as shown in table 2. Further, test compounds, JPC-10 and JPC-13 were found as potent as anticancer compounds tested among the synthesized compounds on EAC cells.

Table 2: Antiglycation and anticancer activity\*

Compound code	IC <sub>50</sub> (µg/ml)		
	BSA-Glucose assay	Hemoglobin-glucose assay	Anticancer activity
JPC-1	-	-	-
JPC-2	-	-	-
JPC-3	340	344.7	-
JPC-4	685	53.3	371.35
JPC-5	736	146	151.50
JPC-6	477	97.3	-
JPC-7	-	724.7	132.49
JPC-8	30	>1000	371.35
JPC-9	600	710.7	191.94
JPC-10	170	56.3	43.50
JPC-11	-	106.9	128.22
JPC-12	229	414.8	67.44
JPC-13	>1000	354.5	-
JPC-14	-	-	-
JPC-15	-	-	-
JPC-16	-	-	-
JPC-17	-	-	-
JPC-18	-	-	-
JPC-19	-	-	-
JPC-20	-	-	-
Quercetin	354.00	-	-
α-Tocopherol	-	12.7	-

\* Data for the compounds showing activity are mentioned in the table

**CONCLUSION**

The γ-benzopyrone pharmacophore in the flavonoid ring could be further modified with other smaller hetero aryl substitution such as thiophenyl substitution at α-position. The 3-hydroxyflavones synthesized with thiophenyl ring at α-position showed moderate *in vitro* antiglycation and anticancer activity. Thus, the test compounds could be further studied for their biological activity in diseases where, the glycation process increases during the pathogenesis or progression.

## REFERENCES

- [1] J. Styskal, H. Van Remmen, A. Richardson, A. B. Salmon. *Free Radic. Biol. Med.*, **2012**, 52, 1, 46.
- [2] V. T. Samuel, G. I. Shulman, *Cell*, **2012**, 148, 5, 852.
- [3] E. Nicolle, F. Souard, P. Faure, A. Boumendjel, *Curr. Med. Chem.*, **2011**, 18, 17, 2661.
- [4] O. Firuzi, R. Miri, M. Tavakkoli, L. Saso, *Curr. Med. Chem.*, **2011**, 18, 25, 3871.
- [5] C. H. Wu, G. C. Yen, *J. Agric. Food Chem.*, **2005**, 53, 8, 3167.
- [6] V. Krystof, S. Uldrijan, *Curr. Drug Targets*, **2010**, 11, 3, 291.
- [7] B. H. Havsteen. *Pharmacol. Ther.*, **2002**, 96, 2-3, 67.
- [8] B. S. Jayashree, A. Alam, Y. Nayak, D. Vijay Kumar, *Med. Chem. Res.*, **2012**, 21, 8, 1991.
- [9] B. S. Jayashree, B. K. Kuppast, K. N. Venugopala, *Asian J. Chem.*, **2007**, 19, 1415.
- [10] B. S. Jayashree, J. C. Thejaswini, Y. Nayak, D. Vijay Kumar, *Asian J. Chem.*, **2010**, 22, 2, 1055.
- [11] B. L. Somani, R. Sinha, M. M. Gupta, *Clin. Chem.*, **1989**, 35, 3, 497.
- [12] S. C. Shivhare, A. O. Patidar, K. G. Malviya, K. K. Shivhare-Malviya, *Ayu.*, **2011**, 32, 3, 388.