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Antimicrobial Activity of 7, 3[°], 4[°]-Trihydroxy flavonol Isolated from *Acacia Nilotica* Var. *Adstringens*

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ABSTRACT

Acacia species (Mimosaceae) is widely distributed in tropical and subtropical countries and has a variety of ethnomedicinal uses. There is inadequate laboratory investigation to identify bioactive compounds and therapeutic effect of Acacia nilotica var. adstringens. This research was conducted to extract, isolate and identify major compounds from heartwood of Acacia nilotica var. adstringens and to test them against representative bacteria. Powdered air-dried heartwood of A. nilotica var. adstringens was extracted with methanol/water, 4:1 and the extracted was then purified using chromatographic techniques (column and paper chromatography). A pure flavonoid compound was isolated and the structure was elucidated based on extensive spectroscopic analysis procedures (IR, UV, 1H-NMR, and MS spectrometery). The isolated compound was then evaluated for antimicrobial potential against Gram-negative (Escherichia coli and Pseudomonas aeruginosa) and Gram-positive bacteria (Bacillus subtilis, Bacillus cereus, and Staphylococcus aureus) using cub-plate agar diffusion method. The spectroscopic analysis of the isolated compound led to its identification as 7, 3`, 4`-trihydroxyflavonol. The compound showed varying antimicrobial responses with high potency against Gram-negative human pathogens. The Gram-positive bacteria which were inhibited were Bacillus cereus, Corynebacterium sp., Enterococcus faecalis, Staphylococcus aureus, and Streptococcus agalactiae. The Gram-negative bacteria were Acinetobacter baumannii, Acinetobacter sp., Escherichia coli, Pseudomonas aeruginosa and yeasts were Candida albicans and Cryptococcus neoformans. The present study demonstrated that 7, 3`, 4`-trihydroxyflavonol is an effective antimicrobial compound. If applied in suitable pharmaceutical formulations it could be valuable for treating various bacterial infections or introduced as adjunct treatment along with standard agents.

Keywords: Ethnomedicine, Acacia species, Flavonols, Spectroscopic analysis, In vitro antimicrobial Activity

INTRODUCTION

Acacia nilotica is the most significant genus of family Leguminosae firstly described by Linnaeus in 1773 [1]. This plant is therapeutically used as anti-cancer, antiscorbutic, astringent, anti-oxidant, natriuretic, antispasmodic, diuretic, intestinal pains, and diarrhea, nerve stimulant, colds, congestion, coughs, dysentery, and fever [2]. Pods not necklace-like, margins straight or create and rarely narrowly constricted between the seeds, if constricted then only occasionally along with the pod. Branches and pods densely pubescent to tomentose especially when young. Pods often rather wide (1.3 cm-2.2cm) with distinctly and often irregularly crenate margins, it occurrence in Algeria, Burkina Faso, Cameroon, Cape Verde, Chad, French Guiana, Gambia, Ghana, Guinea Bissau, India, Iran, Ivory Coast, Libya, Mali, Niger, Nigeria, Oman, Pakistan, Senegal, Somalia, Sudan,

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and Togo [3,4]. It uses as an effective medicine in the treatment of malaria, sore throat (aerial part) and toothache (bark) [5].

The powdered bark of the plant with a little salt is used for treating acute diarrhea. 6 The gum or bark is used for cancer of ear, eye, or testicles, induration of the liver, spleen and excess flesh. Bark, leaves, and young pods are strongly astringent due to tannins and are chewed in Senegal as antiscorbutic, park decoction drunk for intestinal pains and diarrhea. Other preparations are used for coughs, toothache, ophthalmia, and syphilitic ulcers. In Lebanon, the resin is mixed with orange-flower and used as an infusion for typhoid convalescence [6,7] Egyptian Nubians believe that diabetics may eat unlimited carbohydrates as long as they also consume powdered pods. 7 Extracts of the plant were found to be inhibitory to at least four species of pathogenic [8].

Flavonoids are natural phenolic compounds which appear as secondary metabolites of the plant, [9] they are found in many plant tissues, which present inside of the cells or on the surfaces of different plant organs. The flavonoids may be modified by hydroxylation, methylation, or O-glycosylation of hydroxyl groups as well as C-glycosylation directly to the carbon atom of the flavonoid skeleton. Also, alkyl groups (or phenyls) may be covalently attached to the flavonoid moieties, and sometimes additional rings are condensed to the basic skeleton of the flavonoid core [10]. Flavonoids occur both in the (Free State) and as glycosides; their chemical structure is based on a C15 skeleton consisting of two benzene rings connected by a three-carbon chain, that is, C6-C3-C6. The three-carbon on chain is generally closed to form a heterocyclic ring (the C-ring) [11]. The position of the substitution also affects the properties. The flavonols containing two ortho or para hydroxyl in the 2-phenyl ring have anti-oxidant properties, while free hydroxyl groups at the 5, 7-positions have a pro-oxidant effect. Apigenin and genkwanin, which are present in the Chinese, drug "Yuen -Hua" are believed to have diuretic and anthelmintic properties [12].

Some flavonoids like myricetin and kaempferol3-glucoside have an anti-HIV-I potency at a non-toxic concentration [13]. Some of the minor flavonoids have very interesting activities. They have anti-microbial, anti-fungal and cytotoxic properties. 10 Anthocyanin, peonin, and isorhamnetin of the algae Chlamydomonas are highly potent sex-determining hormone. 8 The health effects of flavonoids have long been recognized for anti-oxidant, anti-inflammatory, anti-allergic, hepatoprotective, antithrombotic, antiviral, and anti-carcinogenic properties [14]. Isoflavones, the bioactive ingredient in leguminous vegetables, not only cause a small reduction in blood cholesterol but also reduce blood pressure, arterial dimensions, and oxidative stress [15].

RESULTS AND DISCUSSION

Identification of the isolated compound

The IR spectrum of a compound (Figure 1) (KBr disc) spectrum was showed v 3520(OH), 1608 (C=O), 1569, 1528, and 1476 (C=C, Ar), 1116 (C-O), 854, 808, 770 cm⁻¹ (C-H, Ar, bending).

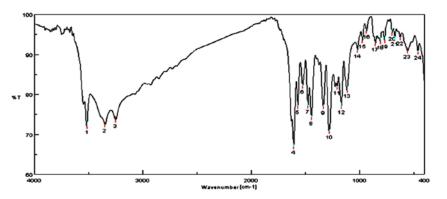


Figure 1: IR spectrum of the isolated compound.

The isolated compound gave UV absorption characteristic flavonol revealed λ max (MeOH) 248, 363 nm (Figure 2A). Addition NaOMe to a methanolic solution gave a 41 nm (Figure 2B) bathochromic shift in a band I without a decrease in intensity and this is diagnostic of a free 4⁻OH. Sodium acetate spectrum (Figure 2C) gave a 17 nm band II bathochromic shift which is indicating a 7-OH function. The boric acid spectrum (Figure 2D) revealed an 18 nm bathochromic shift in band 1, indicating the presence of a B ring catechol moiety. When AlCl3 was added to a methanolic solution, a band I shifted bathochromically by 67 nm (Figure 2E). This indicates a 3- or 5-OH or an ortho-dihydroxyl system. The spectrum was acid-stable (Figure 2F) this indicating of a 3-OH function.

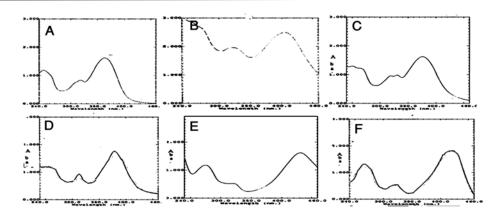


Figure 2: Spectrum of the compound using different solvents, A) methanol; B), sodium methoxide, C), sodium acetate; D), boric acid; E) aluminum chloride and F), Al Cl₃ / HCl.

¹H-NMR showed doublet at δ 7.8 ppm (1H) characteristic of C5- proton (Figure 3). The doublet at δ . 6.8 ppm (2H) is characteristic of C6- and C8protons. The doublet at δ . 7.6 ppm (2H) is account for C2⁻ and C6⁻-protons while the doublet at δ .7.4 ppm (1H) was assigned for C5⁻ proton.

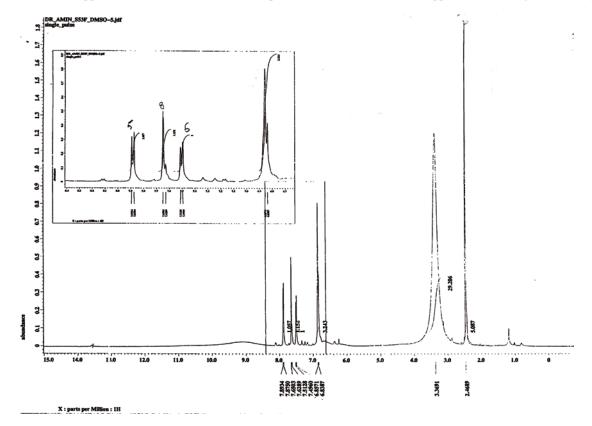


Figure 3: ¹H-NMR spectrum of the isolated compound.

Mass spectrum of the isolated compound shown at base beak m/z 286 (Figure 4). According to the above cumulative data, the following structure suggested a compound is: 7, 3[°], 4[°]-trihydroxyflavonol (Figure 5).

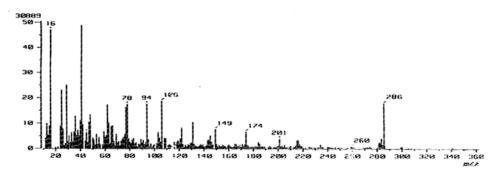
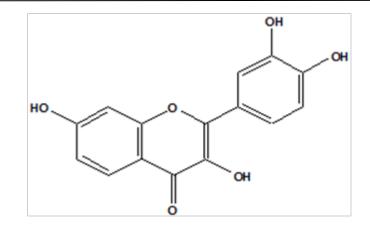
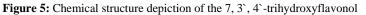


Figure 4: Mass spectrum of the isolated compound.





Antimicrobial activity

The antimicrobial activity of the tested compound was examined against selected gram positive-bacteria and gram-negative bacteria in comparison to reference antibiotics (Figure 6). Results revealed that the tested compound had different antimicrobial responses (Table 1). The gram-positive bacteria which were inhibited were *Bacillus cereus, Corynebacterium* sp., *Enterococcus faecalis, Staphylococcus aureus,* and *Streptococcus agalactiae.* The gram-negative bacteria were *Acinetobacter baumannii, Acinetobacter* sp., *Escherichia coli, Pseudomonas aeruginosa.* The inhibited yeasts were *Candida albicans* and *Cryptococcus neoformans.*

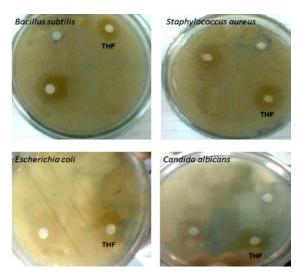


Figure 6: Antimicrobial activities of 7, 3`, 4`-trihydroxyflavonol (THF) isolated from *Acacia Nilotica* var. *adstringens* (inhibition zone diameter mm/mg sample). Abbreviations:1, 1 mg/mL; 1, 10 mg/mL; 3, 100 mg/mL.

 Table 1: Antimicrobial activities of the 7, 3°, 4°-trihydroxyflavonol (THF) isolated from Acacia nilotica var. adstringens (inhibition zone diameter mm /mg sample).

Microorganism	Source / Code	Inhibition zone diameter (mm / mg sample)
Gram-positive bacteria		
1. Bacillus cereus	Laboratory isolates	15
2. Corynebacteriumdiphtheriae	ATCC 13812	10
3. Enterococcus faecalis	ATCC 19433 ^T	12
4. Staphylococcus aureus	ATCC 25923	26
5. Staphylococcus aureus	Clinical isolate	18
6. Streptococcus agalactiae	ATCC 13813 ^T	15

Gram-negative bacteria		
7. Acinetobacter baumannii	<i>ATCC 19606^T</i>	19
8. Acinetobacter sp.	Clinical sample	20
9. Escherichia coli	ATCC 25922	22
10. E. coli	Clinical sample 4662	20
11. Pseudomonas aeruginosa	ATCC 27853	22
Yeasts		
12. Candida albicans	ATCC 10231	15
13. Cryptococcus neoformans	Clinical isolate; VY 4662	16

n conclusion, the compound, 7, 3[,], 4[,]-trihydroxyflavonol, was extracted and isolated from *A. nilotica* var. *adstringes* (collected from Kordofan area, western Sudan). The isolated compound was extracted with methanol and purified using the TLC technique, the structure of the isolated compound was studied and identified using Uv & vis, IR, H1-NMR, and Mass spectra. The biological activity was studied and showed good results.

Plant materials

EXPERIMENTAL

The heartwood of A. nilotica var. adstringens was collected from Elobied, North Kordofan, Sudan. The plant was authenticated by the Department of Botany, University of Khartoum, Sudan.

Extraction of flavonoids

Powdered air-dried heartwood (1 kg) of A. nilotica var. adstringens was extracted with 80% methanol (5 L) at ambient temperature for 72 hours. The solvent was removed in vacuum to give a crude product.`

Isolation method

The methanol extracts of *A.nilotica* var. *adstringens* were separately slurred with water, mixed with a small amount of polyamide and applied on top of the polyamide (800 g) column (100×5 cm). Stepwise gradient elution was carried out using a solvent system of decreasing polarity starting with 100% water and ending with 100% methanol. Fractions of 250 ml were collected and investigated by PC (what man No. 1) using three solvent systems: BAW, 15% AcOH and distilled water. Chromatograms were visualized under UV light before and after exposure to ammonia vapor. Similar fractions were companies and concentrated to dryness under reduced pressure to obtain main fractions. The detected compound was isolated by subsequent PPC from column fraction using different solvent systems (BAW, 15% AcOH). The isolated compound was further purified by rechromatography on Sephadex LH-20 column using MeOH/H₂O, 4: 1.

Preparation of bacterial suspensions

One ml aliquots of 24-hours growth culture of the test organisms were aseptically distributed on to agar slopes and incubated at 37° C, for 24 hours. The bacterial growth was harvested and washed off with sterile normal saline, and finally suspended in 100ml of normal saline to produce a suspension containing about 108–104 colony forming units per ml. The suspension was stored in the refrigerator at 4°C until used. The average number of the viable organism per ml of the saline suspension was determined through the surface viable counting technique. Serial dilution of the stock suspension was made in sterile saline in tubes and on drop volumes (0 ml-20ml) of the appropriate dilution were transferred by adjustable volume micropipette onto the surface of dried nutrient agar plates. The plates were allowed to stand for two hours at room temperature for dry and then incubated at 37°C for 24 hours.

Antimicrobial assay

The cub-plate agar diffusion method adopted with some minor modifications, to assess the antimicrobial activity of Munich bases and their reaction products with o-phenylenediamine, 2 ml of the standardized bacterial stock suspension was mixed with 200 ml of sterile molten nutrient agar which was maintained at 45°C in the water bath. (20 ml) Aliquots of the incubated nutrient agar were distributions into sterile Petri dishes; the agar was left to settle in each of these plates which were divided into two halves. Two cubs in each half (10 mm in diameter) were cut using sterile cork borer (No. 4). Each of the wholes was designed for one of our compounds. Separate Petri dishes were designed for standard antibacterial chemotherapeutic (ampicillin and gentamycin).

The agar discs were removed, a hamates cup were filled with 0.1 ml samples of each compounds using adjustable volume microtiter pipette and allowed to diffuse at room temperature for two hours. The plates were then incubated in the upright position at 37°C for hours.

The above procedure was repeated for different concentrations of the isolated compounds and the standard antimicrobial chemotherapeutic. After incubation, the diameters of resultant growth inhabitation zones were measured.

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REFERENCES

[1] S. Anita, VK Shakya, A Neetu et al., RJPBC. 2012. 3: p. 84-5.

- [2] E. Grotewold. The Science of Flavonoids. New York: Kluwer Academic Pub., 2006.
- [3] RD Barnes and CF Fagg. Invited paper, IUFRO XX World Congress. 6-12 August, 1995, Tampere, Finland. p. 12.
- [4] RK Mishra, M Ramakrishna, V Mishra et al., Current pharmaceutical biotechnology. 2016. 17: 540-8.
- [5] R. Barnes and SA Harris. African Acacias monographs and Manuals Final Technical Report. 2002, p. 21-26.
- [6] Rehman SU, Choe K and Yoo HH. Molecules. 2016; 21: 331.
- [7] S. Amos, PA. Akah, CJ. Odukwe et al, Phytotherapy research PTR, 1999. 13: p. 683-685.
- [8] MB. Sadiq, J. Tarning, TZ Aye Cho et al., Molecules. 2017. 22.
- [9] D. Lin, M. Xiao, J. Zhao et al., Molecules. 2016. 21.
- [10] JA. Duke, P. Duke and JL. DuCellier. Boca Raton, FL: CRC Press, 2008. p. 528.
- [11] A. Atif, A. Naveed, AK. Barkat et al., Journal of Medicinal Plants Research. 2012. 6: p. 1492-1496.
- [12] TA. Akintunde, HM. Babayi and S. Alfa. Nigerian Journal Biotechology. **2015**. 29: p. 34-37.
- [13] AC. Abreu, SC Serra, A. Borges et al. Combinatorial. Microbial drug resistance. 2015. 21: p. 600-609.
- [14] E. Middleton, Jr. Kandaswami and TC. Theoharides. Pharmacological reviews. 2000. 52: p. 673-751.
- [15] K. Taku, K. Umegaki, Y. Sato et al., The American journal of clinical nutrition. 2007. 85: p. 1148-1156.